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(54) Nucleophilic polysubstituted aryl acridinium esters, conjugates, production and uses thereof.

(57) Certain nucleophilic polysubstituted aryl acridinium esters are disclosed, as well as conjugates thereof, which are useful in biological assays, including assays for the determination of vitamin B₁₂, folate, cortisol, estradiol and thromboxane B₂.

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NUCLEOPHILIC POLYSUBSTITUTED ARYL ACRIDINIUM ESTERS, CONJUGATES, PRODUCTION AND USES THEREOF.

This invention relates to nucleophilic polysubstituted aryl acridinium esters and to conjugates, production and uses thereof; more particularly, it relates to assays utilizing the nucleophilic polysubstituted aryl acridinium esters and conjugates thereof.

The use of acridinium esters as chemiluminescent labels in clinical assays is known. For example, European Patent No. 82636 describes the use of an aryl acridinium ester linked to an N-succinimidyl moiety as a chemiluminescent label in immunoassays. U.S. Patent No. 4,745,181 and copending U.S. Patent Application Serial No. 133,792, filed December 14, 1987, describe polysubstituted aryl acridinium esters which are useful in immunoassays and nucleic acid hybridization assays. European Patent Application 89 307 752.9 describes hydrophilic polysubstituted aryl acridinium esters and conjugates thereof useful in clinical assays, particularly those assays involving liposomes.

Richardson et al (Clin. Chem. 31/10, 1664-1668, 1985) and Miller et al (Ann. Clin. Biochem 25, 27-34, 1988) describe the use of (2-aminoethyl)-4-phenyl acridine-9-carboxylate in a chemiluminescent immunoassay for plasma progesterone.

However, the prior art acridinium esters often cannot effectively form conjugates with certain analytes. These analytes may lack nucleophilic groups or may contain carboxylic groups which are not readily amenable to modifications. In some cases, the nucleophilic group of the analyte cannot be acylated with an active-group containing acridinium ester because of the resulting deleterious effect on the immunoactivity of the analyte.

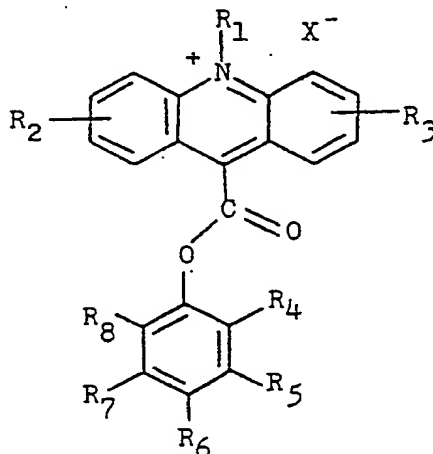
The acridine ester of Richardson et al and Miller et al cannot be converted to a useful acridinium ester without the concurrent loss of the nucleophilicity of the ester. The acridine ester must be conjugated first with the target analyte through a nucleophilic reaction, and then subsequently converted to an acridinium ester moiety. The general reaction conditions of the acridine ester-to-acridinium ester conversion are non-selective. As a result, susceptible groups on the target analytes are frequently affected, resulting in the loss of or reduction of the immunoactivity of the resulting conjugate.

Accordingly, it is an object of the present invention to provide certain advantageous nucleophilic polysubstituted aryl acridinium esters, which may contain an additional ionizable group.

It is a further object of the present invention to provide conjugates formed from the nucleophilic polysubstituted aryl acridinium esters.

It is another object of the present invention to provide assays utilizing the nucleophilic polysubstituted aryl acridinium esters and conjugates thereof.

The present invention provides an acridinium ester characterised in that it corresponds to the following general formula:



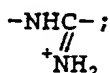
wherein

R₁ represents alkyl, alkenyl, alkynyl, aryl or aralkyl, which may contain one or more heteroatoms; R₂, R₃,

R_5 and R_7 independently represent hydrogen, amino, amido, acyl, alkoxyl, hydroxyl, $-\text{CO}_2$, halide, nitro, $-\text{CN}$, $-\text{SO}_3$, $-\text{NH}-\text{C}(=\text{O})\text{R}$, $-\text{C}(=\text{O})\text{R}$, $-\text{C}(=\text{O})\text{OR}$, $-\text{C}(=\text{O})\text{NHR}$, or $-\text{SCN}$;

wherein R represents alkyl, alkenyl, alkynyl, aryl or aralkyl, which may contain one or more heteroatoms; R_4 and R_8 independently represent hydrogen, alkyl, alkenyl, alkynyl, aralkyl or alkoxyl; R_6 represents $\text{Q}-\text{R}-\text{Nu}$,

$\text{Q}-\text{R}-\text{Nu}$ or $\text{Q}-\text{Nu}$; wherein Q represents $-\text{O}-$, $-\text{S}-$, $-\text{NH}-$, $-\text{C}(=\text{O})-$, $-\text{NH}-\text{C}(=\text{O})\text{NH}-$, $-\text{NH}-\text{C}(=\text{O})\text{NH}-$, $-\text{NH}-\text{C}(=\text{O})\text{O}-$, $-\text{NHC}(=\text{O})-$, $-\text{C}(=\text{O})\text{NH}-$, diazo or

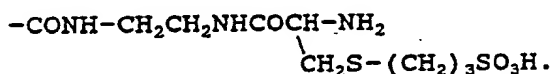


R is as defined above; I represents $-\text{SO}_3$, $-\text{OSO}_3$, $-\text{PO}_3$, $-\text{OPO}_3$ or $-\text{CO}_2$; Nu represents a nucleophilic group; and X represents an anion.

More particularly, it relates to such acridinium esters wherein R_1 represents alkyl, alkenyl, alkynyl or aryl, which may contain up to 24 carbon atoms and/or may contain up to 20, preferably up to 10, heteroatoms, which may be selected from nitrogen, oxygen, phosphorus and sulfur; R_2 , R_3 , R_5 and R_7 independently represent hydrogen, amino, $-\text{CO}_2$, $-\text{CN}$, hydroxyl, C_1-C_4 alkoxyl, nitro, halide, $-\text{SO}_3$ or $-\text{SCN}$; R_4 and R_8 independently represent alkyl, alkenyl, alkynyl or alkoxyl, which may contain up to 8 carbon atoms; X represents halide, CH_3SO_4^- , OSO_2F^- , $\text{OSO}_2\text{CF}_3^-$, $\text{OSO}_2\text{C}_4\text{F}_9^-$, or



R may contain up to 24 carbon atoms and/or may contain up to 20, preferably up to 10, heteroatoms, which may be selected from nitrogen, oxygen, phosphorus and sulfur; and Nu represents amino, hydroxyl, sulfhydryl, active methylene, or an organo-metallic moiety. Preferably, R_1 represents C_1-C_{10} alkyl; R_2 , R_3 , R_5 and R_7 independently represent hydrogen, C_1-C_4 alkoxyl, $-\text{CN}$, $-\text{SO}_3$, nitro or amino; R_4 and R_8 independently represent C_1-C_4 alkyl; and X represents halide. More specifically, R_1 , R_4 and R_8 represent methyl; R_2 , R_3 , R_5 and R_7 represent hydrogen; X represents bromide; and R_6 represents $-\text{CONH}-\text{CH}_2\text{CH}_2-\text{NH}_2$ or



The present invention also provides a luminescent conjugate characterised in that it comprises such an acridinium ester covalently-bound to a biologically-active molecule, such as a protein, an antigen, a hapten, a nucleic acid, or a molecule comprising nucleic acids; preferably vitamin B_{12} or a derivative thereof; or folate or a derivative thereof; or 17-beta-estradiol or a derivative thereof; or cortisol or a derivative thereof; or thromboxane B_2 or a prostaglandin analog.

The present invention further relates generally to the production of such esters and conjugates. Several specific instances may be exemplified. In one embodiment, the present invention provides a process for the production of a conjugate of folate or a derivative thereof and such an acridinium ester characterised in that it comprises;

(a) incubating the folate (derivative) with an amino-reactive compound to couple an acyl or alkoxy carbonyl group to the C-2 amino group of the folate (derivative) so as to form a protected folate intermediate;

(b) activating at least one carboxylic group of the protected folate intermediate;

(c) incubating the activated protected folate intermediate with the acridinium ester so as to form an acridinium ester-protected folate intermediate conjugate; and

(d) incubating the acridinium ester-protected folate intermediate conjugate in an acidic medium to remove the amino-protecting group from the C-2 amino group, so as to form an acridinium ester-folate conjugate. In the case of vitamin B₁₂ or a derivative thereof, the process may comprise:

(a) incubating the vitamin B₁₂ (derivative) in an acidic medium to deaminate the primary propanamide side chains of the Corrin ring of the vitamin B₁₂ (derivative) so as to produce a mixture of carboxylated vitamin B₁₂;

(b) subjecting the mixture of carboxylated vitamin B₁₂ to HPLC to separate the tricarboxylic, the dicarboxylic, and the three monocarboxylic vitamin B₁₂ forms;

(c) isolating one of the monocarboxylic vitamin B₁₂ forms;

(d) activating the carboxylic group of the monocarboxylated vitamin B₁₂; and

(e) incubating the activated monocarboxylic vitamin B₁₂ with the acridinium ester so as to produce an acridinium ester- vitamin B₁₂ conjugate. For 17-beta-estradiol or a derivative thereof, or cortisol or a derivative thereof, the production may comprise:

(a) activating a functional group attached to the 17-beta-estradiol (derivative) or cortisol (derivative) so as to form an activated estradiol or cortisol intermediate; and

(b) incubating the activated estradiol or cortisol intermediate with the acridinium ester so as to form an acridinium ester-17-beta-estradiol or - cortisol conjugate. The production of a conjugate of thromboxane B₂ or a prostaglandin may involve:

(a) activating the terminal carboxylic group of the thromboxane B₂ or prostaglandin analog so as to form an activated intermediate of the thromboxane B₂ or the prostaglandin analog; and

(b) incubating the activated intermediate with the acridinium ester so as to form an acridinium ester-thromboxane B₂ or acridinium ester-prostaglandin analog conjugate.

Furthermore, the present invention relates generally to the use of such acridinium esters and conjugates as labels in assay systems. One method for the determination of vitamin B₁₂ or folate is characterised in that it comprises:

(a) incubating a sample fluid under conditions which will release vitamin B₁₂ or folate in the sample from any endogenous vitamin B₁₂ - or folate - binding proteins so as to produce a released sample fluid;

(b) incubating the released sample fluid with (i) a composite comprising a vitamin B₁₂ - or folate - binding protein immobilized on a solid support and (ii) an appropriate conjugate as above which may be produced by such a process so as to form a complexed composite;

(c) separating the complexed composite from any unbound conjugate;

(d) measuring the amount of acridinium ester label associated with the complexed composite; and

(e) determining the amount of vitamin B₁₂ or folate in the sample fluid from that measurement. More particularly, such a method may comprise:

(a) incubating a sample fluid with (i) a base, preferably sodium hydroxide, and (ii) a sulfhydryl compound, preferably dithiothreitol, so as to produce a released sample fluid;

(b) incubating the released sample fluid with (i) a composite comprising a vitamin B₁₂ - or folate - binding protein immobilized on a solid support and (ii) an appropriate conjugate as above, which may be produced by such a process so as to form a complexed composite;

(c) separating the complexed composite from any unbound conjugate;

(d) incubating the complexed composite with a thiol-reactive compound, preferably ethyl maleimide;

(e) measuring the amount of acridinium ester label associated with the complexed composite; and

(f) determining the amount of vitamin B₁₂ or folate in the sample fluid from that measurement. In the case of cortisol, such an assay may involve:

(a) incubating a sample fluid with (i) a composite comprising an anti-cortisol antibody immobilized on a solid support and (ii) an appropriate conjugate as above, which may be produced by such a process so as to form a complexed composite;

(b) separating the complexed composite from any unbound conjugate;

(c) measuring the amount of acridinium ester label associated with the complexed composite; and

(d) determining the amount of cortisol in the sample fluid from that measurement. For 17-beta-estradiol or thromboxane B₂, the method may comprise:

(a) incubating a sample fluid with (i) an anti-17-beta-estradiol antibody or an anti-thromboxane B₂ antibody and (ii) an appropriate conjugate as above, which may be produced by such a process so as to form a reaction mixture;

(b) incubating the reaction mixture with a composite comprising an antibody capable of binding to anti-17-beta-estradiol or anti-thromboxane B₂ antibody, which antibody is immobilized on a solid support, to

form a complexed composite;

(c) separating the complexed composite from the supernatant;

(d) measuring the amount of acridinium ester label associated with the complexed composite; and

(e) determining the amount of 17-beta-estradiol or thromboxane B₂ in the sample fluid from that

5 measurement.

For the present purposes, Nu is a nucleophilic chemical group which is electron-rich, has an unshared pair of electrons acting as a reactive site and seeks a positively-charged or electron-deficient site on another molecule. As indicated above, examples of useful nucleophilic groups include amino, hydroxyl, 10 sulfhydryl, or an active methylene group, adjacent to a strong electron-withdrawing group. A strong electron-withdrawing group is a chemical group or substituent which strongly attracts electrons and, therefore, intensifies the positive charge of the carbon atom (or carbonium ion) or nullifies the negative charge of the carbon atom (or carbonium ion), to which the group is attached. Examples of strong electron-withdrawing groups include -NO₂, -CN, -SO₃H, -N(R)₃⁺, -S(R)₂⁺, and -NH₃⁺, wherein R is as defined above.

Organic metallic moieties are also useful nucleophilic groups for the purposes of the present invention.

15 An organic metallic moiety is an organic moiety comprising carbon-metal bonds. Examples of organic metallic moieties include Grignard reagents, lithium compounds and phenylsodium.

As indicated, the present invention also relates to conjugates comprising the above-described acridinium esters covalently-bound to various compounds which can covalently bind to the Nu group of the acridinium ester. To form a useful conjugate, the compound must contain at least one functional group 20 capable of binding to the Nu group of the acridinium ester. Preferably, the conjugates are formed from compounds having biological activity, such as proteins, nucleic acids, antigens and haptens.

If Nu is NH₂, for example, examples of suitable compounds for forming the present conjugates include those compounds which contain functional groups capable of binding with -NH₂, such as: (1) carboxylate groups, as in, e.g., folic acid, carboxylated vitamin B₁₂, vitamin B₁₂-hemisuccinate at the ribose moiety, N-hemisuccinates of T₄-methyl ester and T₃ methyl ester, thromboxane B₂, carboxypropyl-theophylline, 25 penicillins, cortisol-3-carboxymethylloxime, estradiol-6-carboxymethylloxime and morphine-6-hemisuccinate; (2) ketone groups, as in, e.g., 3-ketodigoxigenine; (3) aldehyde groups, as in, e.g., digoxin-dialdehyde and bromouridine dialdehyde; (4) halides, as in, e.g., dinitrofluorobenzene and chlorotriazine derivatives of haptens and proteins; (5) active esters, as in, e.g., N-hydroxysuccinimide and imidate derivatives of haptens 30 and proteins; (6) isocyanate and thioisocyanate, as in, e.g., hapten and protein derivatives.

Brief Description of Drawings

35 Fig. 1 is a diagram showing the separation profile of a mixture of carboxylated vitamin B₁₂ (according to Example 5).

Fig. 2 is a diagram showing the separation profile of another mixture of carboxylated vitamin B₁₂ (run 2 of Example 5).

40 Fig. 3 is a diagram showing the separation profile of still another mixture of carboxylated vitamin B₁₂ (run 3 of Example 5).

Fig. 4 is yet another diagram of a separation mixture of carboxylated vitamin B₁₂ (run 4 of Example 5).

45 Fig. 5a is a peak diagram showing peaks in various separation stages of the dicarboxylated vitamin B₁₂ from Figure 2.

Fig. 5b to 5f are peak diagrams showing the analytical HPLC profile of the separated peaks 1 to 5 of Figure 2.

Fig. 6 is a peak diagram showing the analytical HPLC profile of a mixture of monocarboxylated vitamin B₁₂ (from Example 5).

50 Fig. 7 is a diagram showing the tracer activity of several conjugates of B₁₂-ED-DMAE (according to Example 6).

Fig. 8 is a diagram showing a plot of RLU values against the vitamin B₁₂ concentration (in an assay according to Example 12).

55 Fig. 9 is a diagram of the RLU values plotted against vitamin B₁₂ concentration (in an assay according to Example 12).

Fig. 10 is a diagram showing the RLU values plotted against the folate concentration in a folate assay (according to Example 13).

Fig. 11 is a functional diagram of a standard curve illustrating the displacement of tracer bound to

PMP (in cortisol assay according to Example 14).

Fig.12 is a functional diagram showing a standard curve illustrating the displacement of tracer bound to PMP in estradiol assay (according to Example 15).

Fig.13 is a diagram showing a standard curve illustrating the displacement of tracer bound to PMP (in an thromboxane B₂ assay according to Example 16).

If Nu is -SH, examples of suitable compounds will contain functional groups capable of binding with -SH, such as maleimido, dithiopyridino, or olefin as found in, e.g., hapten and protein derivatives.

If Nu is -OH, examples of suitable compounds for forming the conjugates of this invention include those compounds which contain functional groups capable of binding with -OH, such as oxirane, as found in, e.g., hapten and protein derivatives.

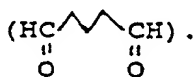
If Nu is a Grignard moiety or other organo-metallic moiety, examples of suitable compounds for forming the conjugates of this invention include those compounds which contain functional groups capable of binding to the moiety, such as ketone and aldehyde, as found in, e.g., aprotic haptens.

It will be appreciated that numerous other suitable Nu groups can be utilized in the acridinium esters of this invention. It is left for the artisan to choose, which combination of acridinium ester and conjugating compound best serves the needs of the desired application.

The term "activation" (or activate) for the purposes of the specification and the claims means a modification of an existing functional group of a particular compound, which modification generates (or introduces) a new reactive functional group from the prior existing functional group, which new reactive functional group is capable of binding to a target functional group of a second compound. For example, the carboxylic group (-COOH) in thromboxane B₂ (see structure below) can be "activated" to produce a mixed anhydride group (-C(=O)-O-C(=O)-R)

using known procedures in the art. The mixed anhydride can then react with the amino group (-NH₂), for example, of 2', 6'-dimethyl-4'-[N-(2 aminoethyl)carbomoyl]phenyl 10-methylacridinium-9-carboxylate bromide (DMAE-ED), to form an amide linkage (-C(=O)-NH-).

resulting in the formation of a Thromboxane B₂-DMAE-ED conjugate (see Example 11 below). As an additional example, the free amino group (-NH₂) group on the surface of alkyl siloxane-coated paramagnetic particles (PMP) (Advanced Magnetics Inc., Cambridge, MA) can be "activated" by derivatization with homobifunctional glutaraldehyde



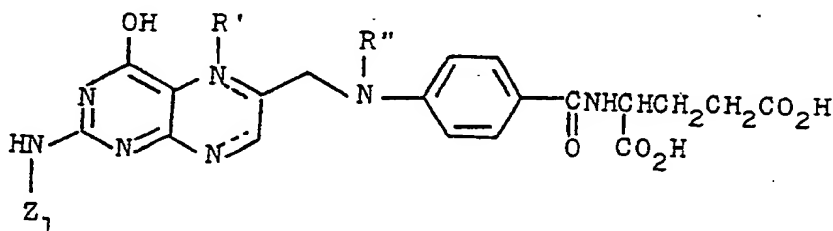
One reactive aldehyde group of the glutaraldehyde covalently binds to the PMP by formation of a Schiff base with the free amino group. The second reactive aldehyde group can then bind with a protein.

The preparation of the conjugates of this invention will vary depending on the acridinium ester and conjugating compound chosen. For example, the following discussion will highlight certain exemplary approaches to forming conjugates from certain compounds and certain preferred Nu groups on the acridinium esters of this invention: (1) when Nu is -NH₂ and the conjugating compound contains a carboxylic group, the carboxylic acid group is first activated to form an active ester, such as N-hydroxysuccinimide ester, mixed anhydride, or acyl halide. The activated compound is then reacted with the acridinium ester to form the conjugate; (2) when Nu is -NH₂ and the conjugating compound contains a ketone or aldehyde group, the acridinium ester can be directly reacted with the compound to form a Schiff base. The conjugate can then be reacted with a hydride reducing agent, such as sodium cyanoborohydride, to stabilize the linkage; (3) when Nu is -NH₂, the acridinium ester can be reacted directly with a conjugating compound containing a reactive group like halide, isocyanate, or thioisocyanate; (4) when Nu is -SH, the conjugating compound should contain a thiol (sulfhydryl)-reactive group, such as maleimido, dithiopyridino, or olefin, to effectively react with the acridinium ester to form a conjugate; (5) when Nu is -OH, it is preferred that the desired conjugating compound contain an oxirane group to effectively react with the acridinium ester to form a conjugate; (6) when Nu is a Grignard or other organo-metallic moiety, the acridinium ester containing such a moiety should be prepared fresh for each use and then reacted with a conjugating compound containing a ketone or aldehyde functional group to form the conjugate.

It will be appreciated that the discussion above is not exhaustive and that numerous other conjugates can be formed from the novel acridinium esters of this invention using known procedures in the art.

The conjugates of this invention are useful as luminescent tracers. The conjugates are particularly useful in luminescent assays using specific binding phenomena such as antibody/antigen immunological reactions, nucleic acid hybridization reactions, or ligand/binding protein interactions.

In one embodiment of the present invention, conjugates are prepared using the acridinium esters of this invention and folate or folate derivatives. Preferably, the acridinium ester used contains both nucleophilic and hydrophilic groups. Preparation of this folate-acridinium ester conjugate involves incubating the acridinium ester with a protected folate intermediate of the following formula (which has been activated at one or both of its carboxylic groups):

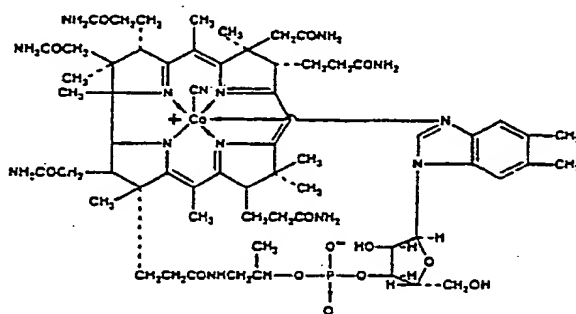


wherein R' is an optional branched or straight-chain, saturated or unsaturated, alkyl of from 1 to 24 carbon atoms, containing 0-20 heteroatoms, preferably nitrogen, oxygen, phosphorous, or sulfur. R'' is Z2, hydrogen, or a branched or straight-chain, saturated or unsaturated, alkyl of from 1 to 24 carbon atoms, containing 0-20 heteroatoms, preferably nitrogen, oxygen, phosphorous, or sulfur. The dotted lines are optional double bonds.

Z1 and Z2 are protecting groups. Z2 is optional. The protecting groups can be any group which can protect the primary and secondary amines from reacting with the activated carboxylic group of the folate either intra- or inter-molecularly. The protecting groups must be removable under conditions which do not deleteriously affect the acridinium ester, preferably in an acidic environment. Useful protecting groups include trifluoroacetyl or t-butyloxy carbonyl groups.

After conjugation of the protected folate intermediate with the acridinium ester, the folate moiety is deblocked by removal of the protecting groups. This deblocking is preferably conducted in an acidic environment using an acidic media such as HBr/acetic acid, which is capable of removing the protecting groups without destroying the integrity of the conjugate. The conjugate so formed can then be used as a tracer in an assay for measuring folates.

In another embodiment of this invention, conjugates are formed using the acridinium esters of this invention and Vitamin B12 (cyanocobalamin). Vitamin B12 has the following structure:



Treating Vitamin B12 with dilute acid will deaminate 1, 2 and/or 3 of the primary propanamide side chains of the Corrin ring to generate a carboxylic function. This carboxylate function is then used to conjugate the Vitamin B12 to the nucleophilic acridinium esters of this invention.

The ratio of monocarboxylic Vitamin B12 (one carboxylate function), dicarboxylic Vitamin B12 (two carboxylate functions), and tricarboxylic Vitamin B12 (three carboxylate functions) will depend on the acid concentration and the reaction time. Monocarboxylic Vitamin B12 is the desired product for the purpose of preparing the conjugates of this invention. Typically, by optimizing known procedures, such as the

procedure described in Allen et al, J. Biol. Chem. 247, 7695 (1972), mixtures of mono-, di-, and tricarboxylic Vitamin B₁₂ can be generated which contain up to 40% monocarboxylic Vitamin B₁₂ (see Figures 3 and 4). Prior art procedures utilize a strong anion exchanger to separate the monocarboxylic Vitamin B₁₂ from the di- and tricarboxylic Vitamin B₁₂. [Allen et al, J. Biol. Chem. 247, 7695, 1972.)

5 There are potentially three forms of monocarboxylic Vitamin B₁₂, depending on which of the 3 primary propanamide side chains have been deaminated. It is desirable to separate these three monocarboxylic Vitamin B₁₂ forms.

It has been unexpectedly discovered that by separating the mixture of carboxylated Vitamin B₁₂ on Reverse Phase High Performance Liquid Chromatography (HPLC), the monocarboxylic Vitamin B₁₂ forms
10 can be separated and isolated from the di- and tri-carboxylic forms and from each other. Accordingly, this allows the artisan to obtain individual monocarboxylic Vitamin B₁₂ forms with high purity without the preliminary ion-exchange fractionation step of the prior art.

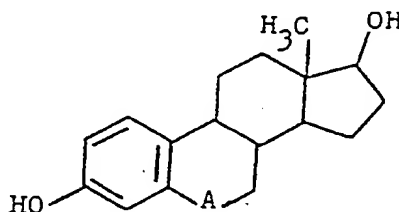
It has also been unexpectedly discovered that one monocarboxylic Vitamin B₁₂ form is more effective than the other two forms in the conjugates of this invention for use in Vitamin B₁₂ assays.

15 In a further embodiment of this invention, conjugates are formed using the acridinium esters of this invention and estradiol. These conjugates can be used, for example, as tracers in assays for 17-beta-estradiol.

17-beta-estradiol has the following structure:

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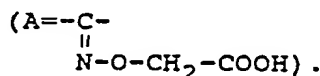
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wherein A is CH₂. Useful derivatives of 17-beta-estradiol include 6-keto-17-beta-estradiol (A = $\text{-}\overset{\text{O}}{\underset{\text{O}}{\text{C}}}\text{-}$)
and, preferably, 6-carboxymethyloxime-17-beta-estradiol

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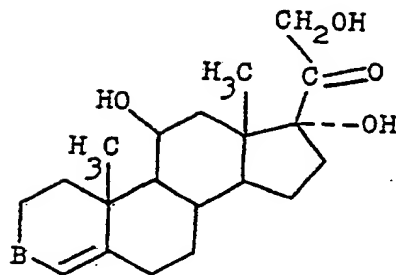


40 Conjugation with an appropriate acridinium ester of this invention can occur at any of the available functional groups on the 17-beta-estradiol or derivative therefore, i.e., the phenolic 3-OH group, the secondary 17-OH group, or the keto or the carboxymethyloxime group created at the C-6 position. The choice of functional group for conjugation will depend generally on such factors as compatibility with the desired immunoassay system, the stability of conjugate prepared, and the ease of preparation. Preferably the conjugate is prepared by activating the carboxylic group of 6-carboxymethyloxime-17-beta-estradiol and
45 then reacting the activated estradiol derivative with an appropriate acridinium ester of the invention. The resulting conjugate can then be used as a tracer in an assay for determining 17-beta-estradiol.

The acridinium esters of this invention can also be used to form useful conjugates with cortisol. Cortisol has the following structure:

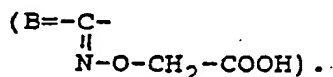
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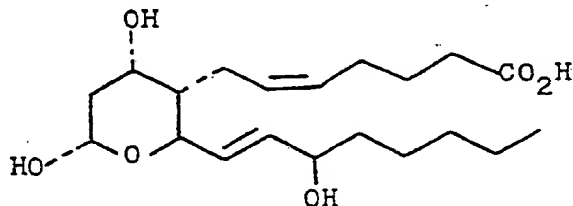
wherein B is - $\text{C}(=\text{O})$ -

A derivative of cortisol is 3-carboxymethyloxime cortisol



Conjugation with an appropriate acridinium ester of this invention can occur at any of the available functional groups on the cortisol or derivative thereof, i.e., the 21-OH group, the 17-OH group, the 11-OH group, the 20-keto group, the 3-keto group, and the 3-carboxymethyloxime group. Preferably, the conjugate is prepared by activating the carboxylic group of the 3-carboxymethyloxime cortisol and then reacting the activated cortisol derivative with an appropriate acridinium ester of this invention. The resulting tracer can then be used as a tracer in an assay for cortisol.

Useful conjugates can be formed between the acridinium esters of this invention and thromboxane B₂ and other prostaglandin analogs. Thromboxane B₂ has the following structure:



Conjugation with an appropriate acridinium ester of this invention can occur at any of the three hydroxyl groups or the olefin functional groups. The choice of functional group will depend generally on such factors as compatibility with a specific binding protein, and low cross reactivity with non-target prostaglandin analogs. Preferably, the conjugate is prepared by activating the terminal carboxylic group of the thromboxane B₂ and then reacting the resulting thromboxane B₂ derivative with an appropriate acridinium ester of this invention. The resulting conjugate can then be used as a tracer in assays for thromboxane B₂.

This invention also relates to assays utilizing the conjugates of this invention as chemiluminescent tracer compounds. The assays can be homogeneous or heterogeneous. The assays can be competitive inhibition assays where, for example, the analyte to be determined is a univalent hapten or molecule. The assays can also be non-competitive, such as sandwich assays where, for example, the acridinium esters of this invention are conjugated to an antibody or a receptor molecule. The components or reagents of the assays utilizing the conjugates of this invention can be mixed together in any desired manner or sequence provided that the resultant acridinium ester label can be measured in a subsequent detection system. Accordingly, the assays utilizing the conjugates of this invention can be conducted in a forward mode, reverse mode, or a simultaneous mode (see, e.g., U.S. Patents Nos. 4,098,876 and 4,244,940).

Assays for the detection and measurement of Vitamin B₁₂ and folate are illustrative of the assays which can be conducted using the conjugates of this invention. Such assays can use the Vitamin B₁₂-acridinium ester or the folate-acridinium ester conjugates of this invention. A general discussion of radioisotope dilution assays for Vitamin B₁₂ and for folate is found in U.S. Patent No. 4,451,571, herein incorporated by

reference.

Assays for the detection or measurement of Vitamin B₁₂ or folate in a sample generally require a sample preparation step wherein the Vitamin B₁₂ or folate in the sample is released (liberated) from endogenous binding proteins. Methods to release the Vitamin B₁₂ or folate from the respective binding proteins include heating or boiling the sample, or using a chemical releasing agent. Typical releasing agents comprise a strong base, such as NaOH. A sulfhydryl compound, such as dithiothreitol (DTT) or beta-mercaptoethanol, is also typically added during the sample preparation step. The sulfhydryl compound can be added before, after, or along with, the addition of the releasing agent.

In one assay for Vitamin B₁₂, following the sample preparation step, the Vitamin B₁₂ tracer compound is combined with the sample and purified hog intrinsic factor (HIF) immobilized on a solid phase. The sample and tracer compound compete for binding sites on the HIF. The amount of tracer compound bound to HIF is inversely proportional to the amount of Vitamin B₁₂ in the sample.

In one assay for folate, following the sample preparation step, the folate tracer compound is combined with the sample and bovine lactoglobulin or folate binding protein (FBP), immobilized on a solid phase. The sample and tracer compound compete for binding sites on the FBP. The amount of tracer compound bound to FBP is inversely proportional to the amount of folate in the sample.

It has been discovered that the sulfhydryl compounds used in the sample preparation step remain in the solid phase at the time of counting (i.e., measuring the amount of label associated with the solid phase). The presence of varying concentrations of the sulfhydryl compound (particularly DTT) in the solid phase can quench the photon output of the chemiluminescent reaction of the acridinium ester label and result in poor assay precision and reduced signal. It was unexpectedly discovered that by incubating the solid phase in a solution comprising a thiol-reactive compound, such as ethyl maleimide, prior to counting, the quenching effect of the sulfhydryl compound is reduced or eliminated. Preferably, the concentration of the thiol-reactive compound in the solution is about 0.01mM to about 50mM, more preferably about 0.5mM to about 10mM, and most preferably, about 1mM.

The following Examples illustrate the present invention.

Example 1

Preparation of 2', 6'-Dimethyl-4'-[N-(2-aminoethyl)carbomoyl]phenyl 10-methylacridinium-9-carboxylate bromide (DMAE-ED)

A solution of 2', 6'-dimethyl-4'-carboxylphenyl 10-methylacridinium-9-carboxylate bromide (DMAE, 200mg, 0.43mmole) (see copending U.S. Application Serial No. 226,639, filed on August 1, 1988) in 30ml of dimethylformamide (DMF) was cooled in ice bath, treated with triethylamine (0.25ml, 1.72mmole), ethylchloroformate (0.08ml, 0.85mmole) and 30ml of chloroform to form a reaction mixture. After 20 min. of stirring, the reaction mixture was transferred to a dried dropping funnel and added dropwise over a 15 minute period to a solution of ethylenediamine in 10ml of DMF/CHCl₃ (1:1) to form a second reaction mixture.

The second reaction mixture was then stirred at room temperature overnight and then evaporated to dryness under vacuo. The residue produced from the evaporation was taken up in 3-4ml of chloroform/methanol/water (65:25:4), purified on two 20x20cm preparative thin layer chromatography TLC plates (Silica gel 60, F254, Merck & Co., Inc., Rahway, NJ) and developed with the same solvent system. The yellow major band which developed (R_f = 0.47) (which could also be detected under long and short UV light) was stripped and eluted with the same solvent system. The eluent was then evaporated. The residue from this evaporation was triturated with 30ml of 10% methanol/chloroform and filtered through Whatman #1 filter paper under gravity. The filtrate so produced was evaporated to produce DMAE-ED (110mg, 50%). Fast Atom Bombardment (FAB) Mass Spectral Analysis (performed by Oneida Research Services, Whitesboro, N.Y.) in the positive ion mode gave a M⁺ peak of 428. Isotopic bromide peaks 79 and 81 of about equal intensity were detected in the negative ion mode.

Example 2

Preparation of N-tert-Butyloxycarbonyl-S-(3-sulfopropyl)cysteine (BOC-SulfoCys)

2-(tert-butoxycarbonyloxyimino)-2-phenylacetoneitrile (BOC-ON) (121mg, 0.49mmole) (Aldrich Chem. Co., Milwaukee, Wisconsin) was added to a solution of S-(3-sulfopropyl)cysteine (SulfoCys, 100 mg, 0.41 mmole), (prepared by the method of U.T. Ruegg and J. Rudinger, J. Peptide Protein Res. 6, 447, 1974), and triethylamine (0.18ml, 1.23mmole) in 3ml of 50% aqueous dioxane to form a reaction mixture. The reaction mixture was heated at 50°-55° C for 1 hour to obtain a yellow solution. The reaction mixture was then cooled, diluted with 5ml of water, and washed with ethylacetate (3x10ml). The resultant aqueous layer was evaporated to dryness under vacuo by coevaporating with methanol twice. TLC (Silica gel 60, Merck & Co., Inc.) analysis of the residue produced by the evaporation, using a solvent system of chloroform/methanol/water (55:40:5), showed complete conversion of SulfoCys to BOC-SulfoCys.

Example 3Preparation of 2', 6'-Dimethyl-4'-[N-[N-(2-tertbutyloxycarbonylamino-3-S-(3'-sulfopropyl)-thiopropionyl)-2-amino ethyl]carbomoyl]phenyl 10-methylacridinium-9-carboxylate bromide (BOC-SulfoCys-ED-DMAE)

A solution of BOC-SulfoCys (3.42mmole) (Example 2), and DMAE-ED (400mg, 0.79mmole) (Example 1) in 110ml of DMF/CHCl₃ (1:1) was treated with dicyclohexylcarbodiimide (325mg, 1.57mmole), stirred at room temperature for 3 hours and evaporated to dryness. The residue from the evaporation was taken up in about 15ml of chloroform/methanol/water (65:25:4) and purified on 8 preparative TLC plates (Silica gel 60, Merck & Co., Inc.) developed with the same solvent system.

The major yellow band which developed at about R_f of 0.55 was stripped and eluted with the same solvent system. The eluent was then evaporated to dryness under vacuo to produce BOC-SulfoCys-ED-DMAE (630mg, 90%)

Example 4Preparation of 2', 6'-Dimethyl-4'-[N-[N-(2-amino-3-S-(3'-sulfopropyl)-thiopropionyl)-2-aminoethyl]-carbomoyl]phenyl 10-methylacridinium-9-carboxylate bromide (SulfoCys-ED-DMAE)

A solution of BOC-SulfoCys-ED-DMAE (630mg, 0.715mmole) (Example 3) in 4ml of 36% HBr/Acetic acid was kept at room temperature for 5 hours to form a reaction mixture. The reaction mixture was added dropwise to about 30ml of anhydrous ethylether, forming a gummy precipitate and supernatant. The supernatant was removed from the precipitate. The precipitate was then dissolved in about 10ml of methanol and the resultant solution was then added dropwise to about 30ml of fresh anhydrous ethylether, forming a yellow precipitate and supernatant. The yellow precipitate and supernatant were then filtered through a medium porosity frit and the resultant yellow solid residue was then washed with anhydrous ethylether, and then air dried to produce SulfoCys-ED-DMAE (438mg, 93.7%).

FAB Mass Spectral analysis (performed by both Oneida Research Services, Whitesboro, N.Y. and Institute of Chemical Analysis, Northeastern University, Boston, MA) in the positive ion mode gave a M⁺ peak of 653.

Example 5Preparation of Monocarboxylic Vitamin B₁₂

A. Preparation of Deaminated Vitamin B₁₂

Vitamin B₁₂ (1.0g, 0.738mmole) (Sigma Chemicals, St. Louis, MO) was added to 80ml of 0.5N HCl to form a reaction mixture and stirred at room temperature for 65 hours. The reaction mixture was then loaded onto a 4x15cm column of Bio-Rad AG1-X8 (acetate form) (Bio-Rad Laboratories, Richmond, CA), 100-200 mesh, packed and eluted as described in Allen, R.H. and Majerus, P.W., J. Biological Chem., 247, 7695-7701 (1972). The initial 300ml of red eluent was collected and evaporated to dryness under vacuum to produce about 1 gram of a mixture of mono-, di-, and tricarboxylated Vitamin B₁₂.

B. Preparation of Mixture of Mono-carboxylated Vitamin B₁₂

The mixture of carboxylated Vitamin B₁₂ prepared in A was fractionated by QAE-Sephadex A-25 chromatography to obtain a mixture of monocarboxylated Vitamin B₁₂ as described by Allen et al. (J. Biol. Chem. 247, 7695, 1972).

C. Preparative HPLC for the Separation of the Carboxylated Vitamin B₁₂

50mg of the mixture of the mono-, di-, and tri-carboxylated Vitamin B₁₂ from A in 2ml of water was injected into a Waters Delta-Prep 3000 HPLC system (Waters Associates, Milford, MA) with an ISCO-Foxy fraction collector, an ISCO-2150 peak separator (ISCO, Lincoln, Nebraska) and a YMC AP 363-15, 30mm x 25cm stainless steel column packed with C18-bonded silica of 15um particles, spherical shape, and 300A pore size (YMC, Inc., Morris Plains, N.J.).

The carboxylated Vitamin B₁₂ was eluted from the column for each run using acetonitrile as Solvent B and 0.05M triethylammonium acetate, pH 4.5, as Solvent A, in the following manner:

1. Run 1 - step-gradient elution:

20 min. on 8% Solvent B/92% Solvent A then
10 min. on 10% Solvent B/90% Solvent A then
50 min. on 15% Solvent B/85% Solvent A

2. Run 2 - isocratic elution:

Using 15% Solvent B/85% Solvent A

3. Run 3 - step-gradient elution:

10 min. on 10% Solvent B/90% Solvent A
10 min. on 15% Solvent B/85% Solvent A

4. Run 4 - isocratic elution:

10% Solvent B/90% Solvent A The flow rate of the column was 20ml/min. for each run and the eluted materials were detected at a wavelength of 280nm.

Figure 1 is a profile of the separation of the mixture of carboxylated Vitamin B₁₂ obtained in Run 1. Five peaks were collected although Peak 5 was not recorded because the preprogrammed recording time ended prior to elution of Peak 5. Peak 5 was determined based on the characteristic red color of the fraction collected. Peak 5 was then added to the graph as a dotted line peak indicating its location had it been recorded.

Figures 2-4 are profiles of the separation of the mixture of carboxylated Vitamin B₁₂ obtained in Runs 2, 3, and 4, respectively.

These results demonstrate that with a properly chosen solvent system and preparative HPLC column, the separation profile of the mixture of the carboxylated Vitamin B₁₂ can be adjusted. For example, as seen in Figure 2 and Figure 5A which show the profile of the analytical HPLC of the same mixture of carboxylated Vitamin B₁₂ (See D below), Peak 3 of fig. 5A (retention time of 13.84 min.) was split into peaks 3 and 4 (Fig. 2) using the preparative column.

D. Analytical HPLC Profile of the Carboxylated Vitamin B₁₂ Derivatives

5 to 20ug of the mixtures of carboxylated Vitamin B₁₂ prepared in A, B, and Run 2 of C, in 20ul of water were each injected into a Beckman 344 HPLC system (Beckman Instruments, San Ramon, CA). The HPLC system contained a 3.9mm x 30cm stainless steel column packed with uBondapak C18 10um particles with irregular shape and 120A pore size (Waters Associates, Milford, MA). The mixtures were each

eluted isocratically from the column using 10% acetonitrile and 90% 0.05M triethylammonium acetate, pH4.5. The flow rate of the elution was 1.5ml/min. and the eluted materials were detected at a wavelength of 280nm.

Figure 5A is a profile of the separation of the carboxylated Vitamin B₁₂ from A. Peak 1 represents the unreacted Vitamin B₁₂, Peak 2 (retention time of 11.97 min.) and peak 3 (retention time of 13.84 min.) represent the 3 incompletely separated monocarboxylated Vitamin B₁₂ forms present in the mixture. Peak 5 in Figure 5A probably represents the dicarboxylated Vitamin B₁₂ form.

Figures 5B-5F are the analytical HPLC profiles of the separated peaks 1-5 (Figure 2) obtained in Run 2 of C. Figure 5D and Figure 5E show the close retention times (13.54 min. and 14.33 min., respectively) of the two different monocarboxylated Vitamin B₁₂ forms which comprised peak 3 of Figure 5A and peaks 3 and 4 of Figure 2.

Figure 6 is the analytical HPLC profile of the mixture of the 3 monocarboxylated Vitamin B₁₂ forms from B. Peaks 1 and 2 of Figure 6 have nearly the same retention times as Peaks 2 and 3 of Figure 5A.

Example 6

Preparation of Conjugates from Monocarboxylic Vitamin B₁₂ and DMAE-ED (B₁₂-ED-DMAE)

Using the 3 monocarboxylic Vitamin B₁₂ forms prepared and isolated in Example 5 (designated hereinafter as monocarboxylic Vitamin B₁₂ forms I, II, and III, which have retention times of 11.58, 13.54, and 14.33 min., respectively) and DMAE-ED prepared in Example 1, three separate but similar conjugating reactions were carried out to prepare the tracers as following:

A solution of monocarboxylic Vitamin B₁₂ form I (10mg., 7.4umole) in 1.8ml of DMF was cooled in ice bath, treated with triethylamine (10.5ul, 74umole, in 100ul DMF) and ethyl chloroformate (2.8ul, 30umole, in 100ul DMF) to form a reaction mixture. After stirring for 30 minutes, the reaction mixture was evaporated to dryness to remove the excess ethyl chloroformate to produce a residue. DMAE-ED (3.4mg, 6.7umole) and triethylamine (5.2ul, 37umole) in 2ml of DMF, were added to the residue to form a second reaction mixture. The second reaction mixture was stirred at room temperature overnight and then evaporated to dryness under vacuo. The crude products so obtained were purified on one analytical silica gel 20x20cm TLC plate (Silica gel 60, Merck & Co, Inc.), developed with chloroform/methanol/water (55:40:5).

Two red bands (hereinafter referred to as the upper and the lower bands) which developed between Rf of 0.47-0.57 were each separately stripped and eluted with the same solvent system. Each eluent was evaporated to dryness to produce a B₁₂-ED-DMAE tracer.

The same procedure described above was repeated for monocarboxylic Vitamin B₁₂ forms II and III. As a result, from the 3 monocarboxylic Vitamin B₁₂ forms, a total of six B₁₂-ED-DMAE conjugates (designated 1 through 6) were isolated. Conjugates 1 and 2 were prepared from form I, 3 and 4 were prepared from form II, and 5 and 6 were prepared from form III. The conjugates were each diluted in phosphate buffered saline (PBS) with 0.1% bovine serum albumin (BSA) and were simultaneously screened for tracer activity (Figure 7) using the following procedure:

A series of Vitamin B₁₂ standards in 1% human serum albumin (HSA) (in 120mM PBS containing 0.2% sodium azide and 0.4g/l merthiolate) were treated by adding 1/20 volume of 1.35M DTT, to produce treated standards. 100ul of each treated standard was added to a 12x75mm plastic tube. To each tube was then added 100ul of 0.5N NaOH and 0.5ml of an IF-PMP (100ug) (prepared as described in Example 12A below except that the suspension contained 3ug IF/g PMP, the heat stress step was omitted, and the IF-PMP was resuspended in 0.16M boric acid, 10mM phosphate, 0.127M NaCl, and 0.1% sodium azide, pH 7.0). 100ul of PBS/BSA containing 12-29x10⁶ relative light units (RLU) (1 RLU = 1 photon count) of the B₁₂-ED-DMAE conjugate to be tested, was added to each tube and the tubes were then incubated at room temperature for one hour. The solid phase in each tube was magnetically separated from the supernatant and the supernatant was then decanted. The solid phase in each tube was washed once with 1ml of water. The solid phase was then resuspended in 100ul water and counted as described in Example 12B below. Table 1 shows the results obtained. In Table 1 T represents the total RLU of each conjugate added, B₀ represents the total RLU associated with the solid phase in the final resuspension, for each conjugate, in the absence of any Vitamin B₁₂, and B₀/T is the percentage of the total RLU added which were associated with the solid phase, for each conjugate.

Table 1

Conjugate	T	B ₀	B ₀ /T %
1	18x10 ⁶	3.8x10 ⁴	0.21
2	29x10 ⁶	5x10 ⁴	0.17
3	17.5x10 ⁶	7.7x10 ⁴	0.44
4	15.2x10 ⁶	6.6x10 ⁴	0.44
5	20.6x10 ⁶	26.1x10 ⁴	1.27
6	12x10 ⁶	20x10 ⁴	1.67

Figure 7 is a plot of B/B₀ against Vitamin B₁₂ concentration for each of the conjugates. B represents the total RLU associated with the solid phase in the final resuspension for a particular concentration of Vitamin B₁₂ in the sample and B₀ is as described above. The conjugate that performed the best (highest B₀/T value) came from the lower band (conjugate #6) which originated from monocarboxylic Vitamin B₁₂ III.

100ug of the B₁₂-ED-DMAE conjugate #6 was separated on an analytical HPLC system as described in Example 5D eluted with mixture of 0.05M triethylammonium acetate, pH 4.5 (Solvent A) and acetonitrile (Solvent B) in linear gradient from 40% B/60% A to 50% B/50% A over a 10 min. period. The flow rate of the eluent was 1 ml/min. and the eluted materials were detected at 260nm. The chromatogram revealed the presence of two peaks (retention times of 5.66 min. and 7.86 min.). These two peaks when isolated and evaluated separately gave identical assay performance whether combined or alone.

Example 7

Preparation of N-Trifluoroacetyl-Folic Acid (TFA-Fol)

A mixture of folic acid (1.0g, 2.27mmole) and trifluoroacetic anhydride (2ml, 6.4mmole) was stirred at room temperature for 1 hour and then evaporated under vacuo. The residue from the evaporation was triturated with a minimal amount of methanol and the supernatant removed by filtration. The resultant wet cake was evaporated to dryness to produce TFA-Fol with an R_f of 0.35 when chromatographed on a TLC plate (Silica gel 60, Merck & Co.) using chloroform/methanol/water (55:40:5).

Example 8

Preparation of Folate-SulfoCys-ED-DMAE conjugate

A solution of TFA-Fol (27mg, 0.05mmole) (Example 7) in 3.75ml of DMF was diluted with 1.8ml of chloroform, cooled in an ice bath and treated with triethylamine (0.065ml, 0.45mmole) and ethyl chloroformate (0.03ml, 0.3mmole) to form a reaction mixture. After 30 min. of stirring, the reaction mixture was evaporated to dryness under vacuo. To the residue which resulted from the evaporation were added 4.5ml of DMF/chloroform (2:1), SulfoCys-ED-DMAE (31mg, 0.04mmole) and triethylamine (0.035ml, 0.24mmole) to form a second reaction mixture. The second reaction mixture was stirred at room temperature overnight and evaporated to form a second residue.

The second residue was purified on a 10x20cm silica gel preparative TLC plate (Silica gel 60, Merck & Co.) developed with chloroform/methanol/water (55:40:5).

The yellow band which developed at about an R_f of 0.64 was stripped and eluted with the same solvent system. The eluent was evaporated to produce crude TFA-Fol-SulfoCys-ED-DMAE (11 mg). TLC analysis of the product showed two major UV positive spots (R_f of 0.6 and 0.5). The spot having R_f of 0.5 was not affected by the subsequent deblocking conditions and was, therefore, considered as an undesirable contaminant.

The crude TFA-Fol-SulfoCys-ED-DMAE obtained above was treated with 200ul of 36% HBr/Acetic acid

at room temperature overnight to form a third reaction mixture. This third reaction mixture was treated with about 10ml of anhydrous ethylether to form a precipitate. After 1. hour of standing, the supernatant was removed from the precipitate by careful pipeting. The precipitate was then dissolved in about 0.5ml of chloroform/methanol/water (65:25:4) and purified on one 20x20cm silica gel analytical TLC plate (Silica gel 60, Merck & Co., Inc.). The TLC plate was developed with chloroform/methanol/water (55:40:5). The yellow band which developed at Rf of 0.38 was stripped and eluted with the same developing solvent system. The eluent was evaporated to produce the Folate-SulfoCys-ED-DMAE conjugate.

Example 9

Preparation of Cortisol-3-CMO-ED-DMAE Conjugate

A solution of 3-carboxymethyloxime-cortisol (Cortisol-3-CMO, 10mg, 0.022mmole) (Steraloids, Wilton, N.H.) in 0.2ml of DMF was diluted with 0.8ml of chloroform, cooled in an ice bath, and treated with dicyclohexylcarbodiimide (DCC, 5.5mg, 0.0266mmole) in 0.2ml of chloroform to produce a reaction mixture. After 10 min. of stirring, the reaction mixture was treated with a solution of DMAE-ED (5.5mg, 0.01mmole) in 0.4ml of DMF to produce a second reaction mixture. The second reaction mixture was stirred at room temperature overnight and evaporated under vacuo. The residue from the evaporation was purified on a 10x20cm silica gel preparative TLC plate (Merck & Co.) and developed with 10% methanol/chloroform. The yellow band which developed at Rf of 0.28 was stripped and eluted with 20% methanol/chloroform. The eluent was evaporated to give Cortisol-3-CMO-ED-DMAE (3.56mg, 42%). FAB Mass Spectral analysis (performed by Institute of Chemical Analysis, Northeastern Univ., Boston, MA) in the positive ion mode gave a M + peak of 845.

Example 10

Preparation of Estradiol-6-CMO-ED-DMAE Conjugate

A solution of 6-carboxymethyloxime-17-beta-estradiol (Estradiol-6-CMO) (23.1mg, 0.062mmole) (Steraloids, Wilton, N.H.) in 2ml of DMF/CHCl₃ (1:1) was cooled in an ice bath, and treated with DCC (15.4mg, 0.074mmole) to produce a reaction mixture. After 10 min. of stirring, the reaction mixture was treated with DMAE-ED (30.1mg, 0.059mmole) to produce a second reaction mixture. The second reaction mixture was stirred at room temperature overnight and evaporated under vacuo. The residue from the evaporation was purified on one 20x20cm silica gel preparative TLC plate developed with 5% methanol/chloroform. The yellow band which developed at Rf of 0.17 was stripped and eluted with 20% methanol/chloroform. The eluent was evaporated to give Estradiol-6-CMO-ED-DMAE (11.9mg, 26%). FAB Mass Spectral analysis (performed by Institute of Chemical Analysis, Northeastern University, Boston, MA) in the positive ion mode gave a M + peak of 789.

Example 11

Preparation of Thromboxane B₂-ED-DMAE Conjugate (TxB₂-ED-DMAE)

A solution of Thromboxane B₂ (TxB₂) (2.5mg, 0.0067mmole) (Biomol, Plymouth Meeting, PA) in 0.4 ml of DMF/CHCl₃ (1:1) was cooled in an ice bath, treated with triethylamine (6ul, 0.04mmole), and ethyl chloroformate (2ul, 0.02mmole) to produce a reaction mixture. After 30 min. of stirring, the reaction mixture was evaporated to dryness. The residue from the evaporation was dissolved in 0.4ml of DMF/CHCl₃ (1:1), treated with triethylamine (6ul, 0.04mmole) and DMAE-ED (4.5mg, 0.009mmole) to produce a second reaction mixture. The second reaction mixture was stirred at room temperature and evaporated under vacuo

to form a second residue. The second residue was taken up in about 0.5ml of chloroform/methanol/water (65:25:4) and purified on a 20x20cm silica gel analytical TLC plate (Merck & Co.) developed with 15% methanol/chloroform.

The major yellow band which developed at Rf of 0.49 was stripped and eluted with 15% methanol/chloroform. The eluent was evaporated to produce TxB₂-ED-DMAE.

Example 12

Vitamin B₁₂ Assay

A. Preparation of Intrinsic Factor Paramagnetic Particles (IF-PMP)

PMP (obtained from Advanced Magnetics Inc., Cambridge, MA) were activated with glutaraldehyde as described in U.S. Patent No. 4,454,083.

To a solution of human serum albumin (HSA) (400mg, Immunosearch, Toms River, N.J.) in 25ml of 10mM sodium phosphate, pH 7.4, was added purified hog Intrinsic Factor (purchased from Dr. R.H. Allen, University of Colorado Medical Center, Denver, CO) (75ug) in 5ml of saline to produce a protein mixture.

The protein mixture was added to a suspension of the activated PMP (5g) in 60ml of 10mM sodium phosphate and shaken at room temperature overnight to produce IF-PMP.

The IF-PMP were then washed and the excessive activated groups were quenched with glycine.

The IF-PMP were resuspended in 200ml of 30mM PBS with 0.1% sodium azide, 0.1% BSA and 0.001% BgG, cured at 50 °C for 16 hrs, washed three times with 10mM sodium phosphate, washed three times with Glycine Buffer (0.325% glycine, 0.1% sodium azide, and 0.1% BSA, pH 7.8), resuspended in the Glycine Buffer (25mg/ml) and stored at 4 °C until needed.

B. Simultaneous Assay

A series of standards in 6% HSA (in 120mM PBS with 0.2% sodium azide and 0.4g/l merthiolate) with known increasing amounts of Vitamin B₁₂, were added to 12x75mm plastic tubes (100ul/tube). 0.1ml of Releasing Agent (0.5M NaOH, 50ug/ml KCN, 0.3ug/ml cobinamide, 0.064M dithiothreitol) was added to the tubes and the tubes were incubated at room temperature for 15 minutes. The IF-PMP prepared in A was diluted 1:312 in the Glycine Buffer (80ug/ml). 0.5ml of the diluted IF-PMP (40ug/tube) and 0.1ml of the B₁₂-ED-DMAE conjugate #6 prepared in Example 6, diluted in PBS with 0.1% BSA and 0.1% sodium azide (4x10⁶ RLU/tube), were then added to the tubes and the tubes were incubated for 60 min. at room temperature.

The tubes were then placed in a magnetic rack useful for magnetic separation of paramagnetic particles in tubes (available from Ciba Corning Diagnostics Corp., Medfield, MA). The magnetic field separated the particles from the supernatant and the supernatant was then decanted. The particles were washed once in 1ml of water, vortexed, magnetically separated from the wash and decanted. The particles were then resuspended in 0.1ml of a 1mM ethyl maleimide.

The tubes were then placed in a luminometer (MAGIC[®] LITE Analyzer, Ciba Corning Diagnostics Corp., Medfield, MA). 0.3ml of a solution of 0.5% hydrogen peroxide in 0.1 N HNO₃ was added to each tube by the luminometer and the light emission was triggered by the addition of 0.3ml of 0.25N NaOH containing ARQUAD surfactant (Armack Chemicals, Chicago, Illinois). The measured RLU's of each tube normalized against the RLU'S of the zero standard were plotted against their respective Vitamin B₁₂ concentrations as shown in Figure 8.

C. Split Incubation Assay

A series of Vitamin B₁₂ standards in 5% HSA (in PBS with 0.2% sodium azide, 2mg/l amphotericin and

24mg/l gentamycin) with known increasing amounts of Vitamin B₁₂ were added to tubes (100ul/tube) and incubated with the Releasing Agent of B except that the cobinamide was omitted. 0.5ml of the diluted IF-PMP of B (40ug/tube), but with 0.06ug/ml cobinamide added to the buffer, was added to each tube and the tubes were then incubated for 45 minutes at room temperature. 0.1ml of the B₁₂-ED-DMAE conjugate #6 (8x10⁵ RLU) prepared in Example 6 diluted with 10mM PBS, pH 7.4, containing 0.1% sodium azide and 0.1% BSA, was then added to each tube and the tubes were then incubated for 30 minutes at room temperature. The particles in the tubes were magnetically separated, washed, resuspended, and counted as described in B. The measured RLU's normalized against the RLU's of the zero standard for each tube were plotted against their respective Vitamin B₁₂ concentration as shown in Figure 9.

Example 13

Folate Assay

A. Reagents

The standards used in the Folate assay were PGA (pteroylglutamic acid) (Sigma Chemical Co, St. Louis, MO) dissolved in 120mM PBS, pH 7.4 with 4% HSA, 0.2% sodium azide, 2mg/l amphotericin, and 24mg/l gentamycin added as preservatives. The folate concentrations were zero, 0.25, 0.5, 1.0, 2.5, 5, 10, 15, 20, and 30ng PGA/ml.

The Releasing Agent was 0.5N NaOH containing 64mM dithiothreitol.

Folate-SulfoCys-ED-DMAE conjugate (8.8x10¹¹ RLU) obtained in Example 8 was first dissolved in 22ml of 10% DMF/water. The solution was then further diluted 1:11250 with 325mM glycine containing 0.1% BSA and 0.1% sodium azide to form a second solution. This second solution was filtered through a 0.2um cellulose acetate filter (Schleicher and Schuell, Keene, N.H.) to produce a tracer solution. 500ul of the tracer solution was added per test.

The binder in the assay was Folate Binding Protein (FBP) (a bovine milk lactoglobulin, purchased from Dr. R.H. Allen, University of Colorado Medical Center, Denver, CO). FBP-PMP and Bovine Gamma Globulin (BgG)-PMP were prepared by the method described in U.S. Patent No. 4,454,088. The FBP-PMP (0.96mg/ml) was diluted 1:60 in 10mM PBS with 0.1% BSA and 0.1% sodium azide, pH 7.4. This was bulked with BgG-PMP at 0.4mg/ml to form the solid phase binder. 100ul of this solid phase binder was added per test, resulting in the addition of 1.6ug FBP-PMP and 40ug BgG PMP. In the final assay there was 100ul of sample or standard, 100ul of Releasing Agent, 500ul of tracer solution and 100ul of solid phase binder, for a total assay volume of 800ul.

B. Assay Procedure

Standards or samples (100ul) were added to 12x75mm polystyrene tubes (Sarstedt, West Germany). To each tube was added 100ul of the Releasing Agent of A. The tubes were vortexed and incubated for 15 min. at room temperature. The tracer solution of A (500ul) was then added to each tube, followed by the addition of the solid phase binder of A (100ul). The tubes were vortexed again and incubated for one hour at room temperature. The tubes were then put on a magnetic separator for 3 minutes, decanted, and blotted. 1ml of deionized water was added to each tube to wash out excess unbound tracer. The solid phase in the tubes was magnetically separated for 3 min., the supernatant decanted, and the tubes drained for 3 minutes. To the resulting pellets in the tubes was added 100ul of 1mM ethyl maleimide. The tubes were then placed in a luminometer and counted as described in Example 12B. The RLU's for each standard were normalized against the RLU's of the zero standard and plotted against the respective folate concentration of the standards to give a displacement curve as shown in Figure 10.

Example 14

Cortisol Assay5 A. Reagent Preparation

The Cortisol-ED-DMAE conjugate of Example 9 was dissolved in methanol and kept at -20° C as a stock solution. The final cortisol-ED-DMAE conjugate was diluted in a buffer containing 10mM sodium phosphate, pH 7.4, 0.1% bovine serum albumin, 0.4mg/ml of 8-anilino-1-naphthalenesulphonate, 0.1% Triton X-100, and 0.05% sodium azide, to produce the tracer solution.

Rabbit anti-cortisol antiserum was bought from Bioclinical Group, Cambridge, MA. The antibody was immobilized on PMP (Advanced Magnetics Inc.) as described in U.S. Patent No. 4,554,088 except that 0.01M sodium acetate buffer, pH 5.5, was used instead of 0.1M sodium phosphate buffer, pH 7.4. The final PMP wet cake was diluted with a buffer containing 0.01M sodium phosphate, 0.1% bovine serum albumin, 4ng/ml 11-deoxycortisol, and 0.4mg/ml 8-anilino-1-naphthalene sulfonate, pH 7.4, to form a PMP suspension (10mg/ml).

20 B. Assay Procedure

25ml each of cortisol standards with concentrations from 0 to 750ng/ml were added to 12x75mm polystyrene test tubes (Sarstedt, West Germany) in duplicate. 100ul of the tracer solution of A with total activity of 10⁵ RLU were then added to the tubes followed by 500ul of the diluted PMP suspension of A. After vortexing, the tubes were incubated for 1 hour at room temperature. The PMP in the tubes were magnetically separated from the supernatant. The supernatant in each tube was then decanted and the PMP in each tube were washed once with 500ul of 0.87% saline and then resuspended in 100ul of water. The tubes were then placed in a luminometer and counted as described in Example 12B. A standard curve in Figure 11 shows the displacement of tracer bound to PMP by added cortisol in the standard. The displacement is inversely proportional to the concentration of the cortisol in the standard.

30 Example 1535 Estradiol Assay40 A. Reagent Preparation

The Estradiol-ED-DMAE conjugate of Example 10 was dissolved in methanol and kept at -20° C as a stock solution. The stock solution was diluted with 0.01M sodium phosphate buffer, pH 7.4, containing 0.1% bovine serum albumin, 0.15M NaCl, and 0.05% sodium azide to form a tracer solution.

45 Monoclonal anti-estradiol antibody was produced in mice (A/J) by immunization with a BSA-estradiol conjugate and subsequent fusion of the splenocytes with Sp2/0-Ag14 myeloma cells by the procedure described by Kohler and Milstein in Nature (London), vol. 256, pp. 495-497 (1975). Hybridoma cells secreting anti-estradiol antibody were detected by the following procedure: Supernatant from the cells were diluted 1:5 in phosphate buffered saline containing 1mg/ml bovine serum albumin. 100ul of each diluted supernatant and 100ul of acridinium ester-labelled estradiol-fowl gamma globulin as tracer were added to a test tube and incubated for one hour at room temperature. Goat anti-mouse IgG coupled to paramagnetic particles were added to each tube and incubated further for 10 minutes at room temperature. The particles were magnetically separated and read on a luminometer for tracer bound to the particles. The cells that tested positive (i.e., produce photon counts over background) were plated at 0.1 cell/well and retested after growth.

55 Cells resulting from this regrowth which tested positive were then injected intraperitoneally into pristane-primed mice (CAF₁). Ascitic fluid from these mice was collected after 3-5 weeks. The anti-estradiol antibody was used directly without further purification.

Goat anti-mouse IgG PMP particles were prepared by immobilizing the IgG fraction of goat anti-mouse IgG antiserum (Jackson Laboratory, PA) on paramagnetic particles by the method described in U.S. Patent No. 4,454,088. The final PMP wet cake was diluted with phosphate buffered saline containing 1mg/ml bovine serum albumin (PBS/BSA) to produce a PMP suspension with a final concentration of 10mg/ml,

B. Assay Procedure

50ul each of a series of estradiol standards with concentration range from 0 to 2000pg/ml were added to 12x75mm polystyrene test tubes (Sarstedt, West Germany) followed by the addition of 100ul of the tracer solution of A with total activity of 385,000 RLU. 100ul of the ascitic fluid from A was diluted 1:20000 in PBS/BSA buffer and then was added to each tube. The tubes were all vortexed and incubated for one hour at room temperature. The PMP suspension of A was diluted in PBS/BSA to a final concentration of 80ug/ml. 500ul of the diluted PMP suspension was then added to each of the test tubes. The tubes were then vortexed and incubated for 30 minutes at room temperature. The PMP in the tubes were then magnetically separated. The PMP in each tube was then washed once with 500ul of saline containing 0.05% Triton X-100, magnetically separated, and the supernatant decanted. The PMP was then resuspended in 100ul of water. The tubes were then placed in a luminometer and counted as described in Example 12B. The displacement curve in Figure 12 shows that the photon counts are inversely proportional to the concentration of estradiol in the standards.

Example 16

Thromboxane B₂ (TxB₂) assay

A. Reagent preparation

The TxB₂-ED-DMAE conjugate of Example 11 was dissolved in methanol and kept at -80 °C as a stock solution. The stock solution was diluted with 0.01M sodium phosphate buffer, pH 7.4, containing 0.1% BSA, 0.15M NaCl and 0.05% sodium azide to produce the tracer solution.

Rabbit anti-TxB₂ antiserum was bought from Cayman Chemicals Co., Ann Arbor, MI. The antiserum was diluted with 0.01M sodium phosphate buffer, pH 7.4, containing 0.15M sodium chloride, 1mg/ml bovine serum albumin, 0.05% sodium azide.

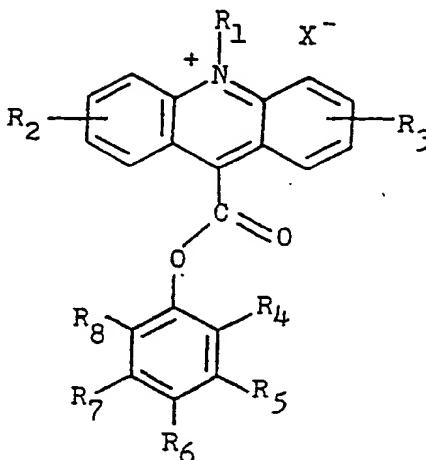
Goat anti-rabbit IgG PMP was prepared by immobilizing the IgG fraction of goat anti-rabbit IgG antiserum (Jackson Laboratory, PA) on paramagnetic particles (PMP) by the method described in U.S. Patent No. 4,454,088. The final PMP wet cake was diluted with PBS/BSA buffer (10mg/ml) to produce a PMP suspension.

B. Assay procedure

100ul of each of a series of TxB₂ standards in PBS/BSA (0-30ng/ml) were added to polystyrene test tubes (12x75mm, Sarstedt, West Germany). 100ul of the tracer solution of A with a total activity of 350,000 RLU, was then added to each tube. 100ul of rabbit anti-TxB₂ antiserum (prepared in A diluted 1/40000 in PBS/BSA) was pipetted into all tubes. All tubes were vortexed and incubated at room temperature for 1 hour. 500ul of the diluted PMP suspension of A was then added to all the tubes and incubated for 45 minutes at room temperature. The PMP in the tubes were then magnetically separated from the supernatant. The supernatant was decanted and the PMP were then washed once with 500ul of water then resuspended in 100ul of water. The tubes were then placed in a luminometer and counted as described in Example 12B. The displacement curve in Figure 13 shows that the photon counts were inversely proportional to the concentration of TxB₂ in the standards.

Claims

1. An acridinium ester characterised in that it corresponds to the following general formula:



wherein

R₁ represents alkyl, alkenyl, alkynyl, aryl or aralkyl, which may contain one or more heteroatoms;
R₂, R₃, R₅ and R₇ independently represent hydrogen, amino, amido, acyl, alkoxy, hydroxyl, -CO₂, halide, nitro, -CN, -SO₃,

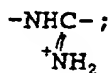
-NH-C(=O)-R, -C(=O)-R, -C(=O)-OR, -C(=O)-NHR,

or -SCN; wherein R represents alkyl, alkenyl, alkynyl, aryl or aralkyl, which may contain one or more heteroatoms;

R₄ and R₈ independently represent hydrogen, alkyl, alkenyl, alkynyl, aralkyl or alkoxy;

R₆ represents Q-R-Nu, Q-R-Nu or Q-Nu; wherein Q represents -O-, -S-, -NH-,
-C(=O)-, -NH-C(=O)-NH-, -NH-C(=O)-NH-, -NH-C(=O)-O-, -NHC(=O)-, -C(=O)-NH-,

diazo or



R is as defined above; I represents -SO₃, -OSO₃, -PO₃, -OPO₃ or -CO₂; Nu represents a nucleophilic group; and

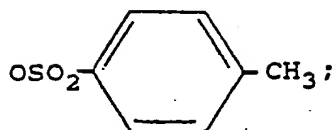
X represents an anion.

2. An acridinium ester as claimed in claim 1 wherein R₁ represents alkyl, alkenyl, alkynyl or aryl, which may contain up to 24 carbon atoms and/or may contain up to 20, preferably up to 10, heteroatoms, which may be selected from nitrogen, oxygen, phosphorus and sulfur;

R₂, R₃, R₅ and R₇ independently represent hydrogen, amino, -CO₂, -CN, hydroxyl, C₁-C₆ alkoxy, nitro, halide, -SO₃ or -SCN;

R₄ and R₈ independently represent alkyl, alkenyl, alkynyl or alkoxy, which may contain up to 8 carbon atoms;

X represents halides CH₃SO₄⁻, OSO₂F⁻, OSO₂CF₃⁻, OSO₂C₄F₉⁻, or



R may contain up to 24 carbon atoms and/or may contain up to 20, preferably up to 10, heteroatoms, which may be selected from nitrogen, oxygen, phosphorus and sulfur;
and

Nu represents amino, hydroxyl, sulfhydryl, active methylene, or an organo-metallic moiety.

3. An acridinium ester as claimed in claim 1 or claim 2 wherein R₁ represents C₁-C₁₀ alkyl;

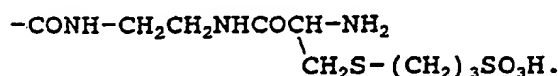
R₂, R₃, R₅ and R₇ independently represent hydrogen, C₁-C₄ alkoxyl, -CN, -SO₃, nitro or amino;

R₄ and R₈ independently represent C₁-C₄ alkyl;

and

X represents halide.

4. An acridinium ester as claimed in any of claims 1 to 3 wherein R₁, R₄ and R₈ represent methyl; R₂, R₃, R₅ and R₇ represent hydrogen; X represents bromide; and R₆ represents -CONH-CH₂CH₂-NH₂ or



5. A luminescent conjugate characterised in that it comprises an acridinium ester as claimed in any of claims 1 to 4 covalently-bound to a biologically-active molecule .

6. A luminescent conjugate as claimed in claim 5 wherein the biologically-active molecule is a protein, an antigen, a hapten, a nucleic acid, or a molecule comprising nucleic acids; preferably vitamin B₁₂ or a derivative thereof; or folate or a derivative thereof; or 17-beta-estradiol or a derivative thereof; or cortisol or a derivative thereof; or thromboxane B₂ or a prostaglandin analog.

7. A process for the production of a conjugate of folate or a derivative thereof and an acridinium ester as claimed in any of claims 1 to 4 characterised in that it comprises:

(a) incubating the folate (derivative) with an amino-reactive compound to couple an acyl or alkoxycarbonyl group to the C-2 amino group of the folate (derivative) so as to form a protected folate intermediate;

(b) activating at least one carboxylic group of the protected folate intermediate;

(c) incubating the activated protected folate intermediate with the acridinium ester so as to form an acridinium ester-protected folate intermediate conjugate; and

(d) incubating the acridinium ester-protected folate intermediate conjugate in an acidic medium to remove the amino-protecting group from the C-2 amino group so as to form an acridinium ester-folate conjugate.

8. A process for the production of a conjugate of vitamin B₁₂ or a derivative thereof and an acridinium ester as claimed in any of claims 1 to 4 characterised in that it comprises:

(a) incubating the vitamin B₁₂ (derivative) in an acidic medium to deaminate the primary propanamide side chains of the Corrin ring of the vitamin B₁₂ (derivative) so as to produce a mixture of carboxylated vitamin B₁₂;

(b) subjecting the mixture of carboxylated vitamin B₁₂ to HPLC to separate the tricarboxylic, the dicarboxylic, and the three monocarboxylic vitamin B₁₂ forms;

(c) isolating one of the monocarboxylic vitamin B₁₂ forms;

(d) activating the carboxylic group of the monocarboxylated vitamin B₁₂; and

(e) incubating the activated monocarboxylic vitamin B₁₂ with the acridinium ester so as to produce an acridinium ester-vitamin B₁₂ conjugate.

9. A process for the production of a conjugate of 17-beta-estradiol or a derivative thereof or cortisol or a derivative thereof and an acridinium ester as claimed in any of claims 1 to 4 characterised in that it comprises:

(a) activating a functional group attached to the 17-beta-estradiol (derivative) or cortisol (derivative) so as to form an activated estradiol or cortisol intermediate; and

(b) incubating the activated estradiol or cortisol intermediate with the acridinium ester so as to form an acridinium ester-17-beta-estradiol or -cortisol conjugate.

10. A process for the production of a conjugate of thromboxane B₂ or a prostaglandin and an acridinium ester as claimed in any of claims 1 to 4 characterised in that it comprises:

(a) activating the terminal carboxylic group of the thromboxane B₂ or prostaglandin analog so as to form an activated intermediate of the thromboxane B₂ or the prostaglandin analog; and

(b) incubating the activated intermediate with the acridinium ester so as to form an acridinium ester-thromboxane B₂ or acridinium ester-prostaglandin analog conjugate.

11. The use of an acridinium ester as claimed in any of claims 1 to 4 or a conjugate as claimed in claim 5 or claim 6 or produced by a process as claimed in any of claims 7 to 10 as a label in an assay system.

12. A method for the determination of vitamin B₁₂ or folate characterised in that it comprises:

(a) incubating a sample fluid under conditions which will release vitamin B₁₂ or folate in the sample from any endogenous vitamin B₁₂ - or folate - binding proteins so as to produce a released sample fluid;

(b) incubating the released sample fluid with (i) a composite comprising a vitamin B₁₂ - or folate - binding protein immobilized on a solid support and (ii) a conjugate as claimed in claim 6 or produced by a process as claimed in claim 8 or claim 7 so as to form a complexed composite;

(c) separating the complexed composite from any unbound conjugate;

(d) measuring the amount of acridinium ester label associated with the complexed composite; and

(e) determining the amount of vitamin B₁₂ or folate in the sample fluid from that measurement.

13. A method for the determination of vitamin B₁₂ or folate characterised in that it comprises:

(a) incubating a sample fluid with (i) a base, preferably sodium hydroxide, and (ii) a sulfhydryl compound, preferably dithiothreitol, so as to produce a released sample fluid;

(b) incubating the released sample fluid with (i) a composite comprising a vitamin B₁₂ - or folate - binding protein immobilized on a solid support and (ii) a conjugate as claimed in claim 6 or produced by a process as claimed in claim 8 or claim 7 so as to form a complexed composite;

(c) separating the complexed composite from any unbound conjugate;

(d) incubating the complexed composite with a thiol-reactive compound, preferably ethyl maleimide;

(e) measuring the amount of acridinium ester label associated with the complexed composite; and

(f) determining the amount of vitamin B₁₂ or folate in the sample fluid from that measurement.

14. A method for the determination of cortisol characterised in that it comprises:

(a) incubating a sample fluid with (i) a composite comprising an anti-cortisol antibody immobilized on a solid support and (ii) a conjugate as claimed in claim 6 or produced by a process as claimed in claim 10 so as to form a complexed composite;

(b) separating the complexed composite from any unbound conjugate;

(c) measuring the amount of acridinium ester label associated with the complexed composite; and

(d) determining the amount of cortisol in the sample fluid from that measurement.

15. A method for the determination of 17-beta-estradiol or thromboxane B₂ characterised in that it comprises:

(a) incubating a sample fluid with (i) an anti-17-beta-estradiol antibody or an anti-thromboxane B₂ antibody and (ii) a conjugate as claimed in claim 6 or produced by a process as claimed in claim 9 or claim 11 so as to form a reaction mixture;

(b) incubating the reaction mixture with a composite comprising an antibody capable of binding to anti-17-beta-estradiol or anti-thromboxane B₂ antibody, which antibody is immobilized on a solid support, to form a complexed composite;

(c) separating the complexed composite from the supernatant;

(d) measuring the amount of acridinium ester label associated with the complexed composite; and

(e) determining the amount of 17-beta-estradiol or thromboxane B₂ in the sample fluid from that measurement.

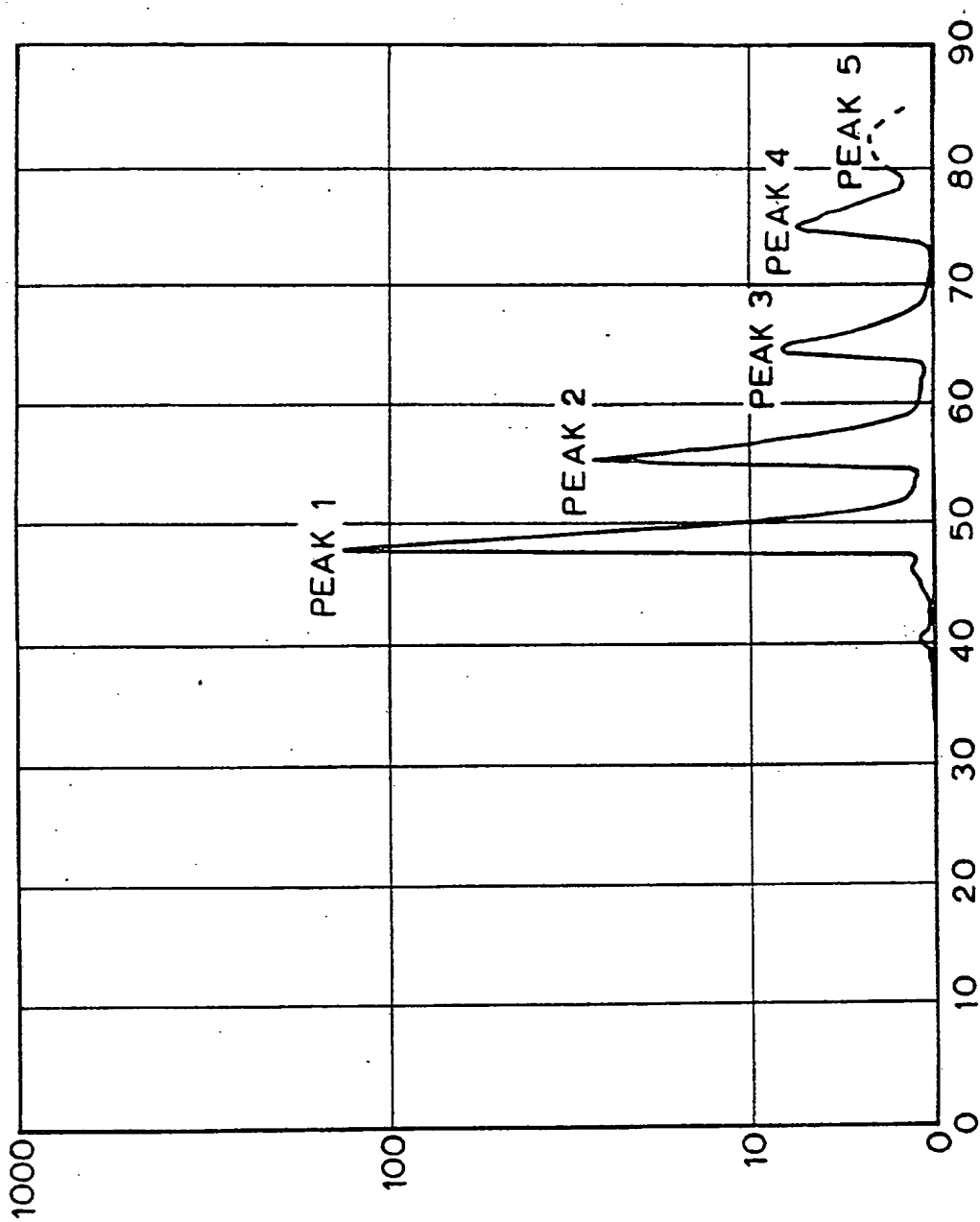


FIG. 1

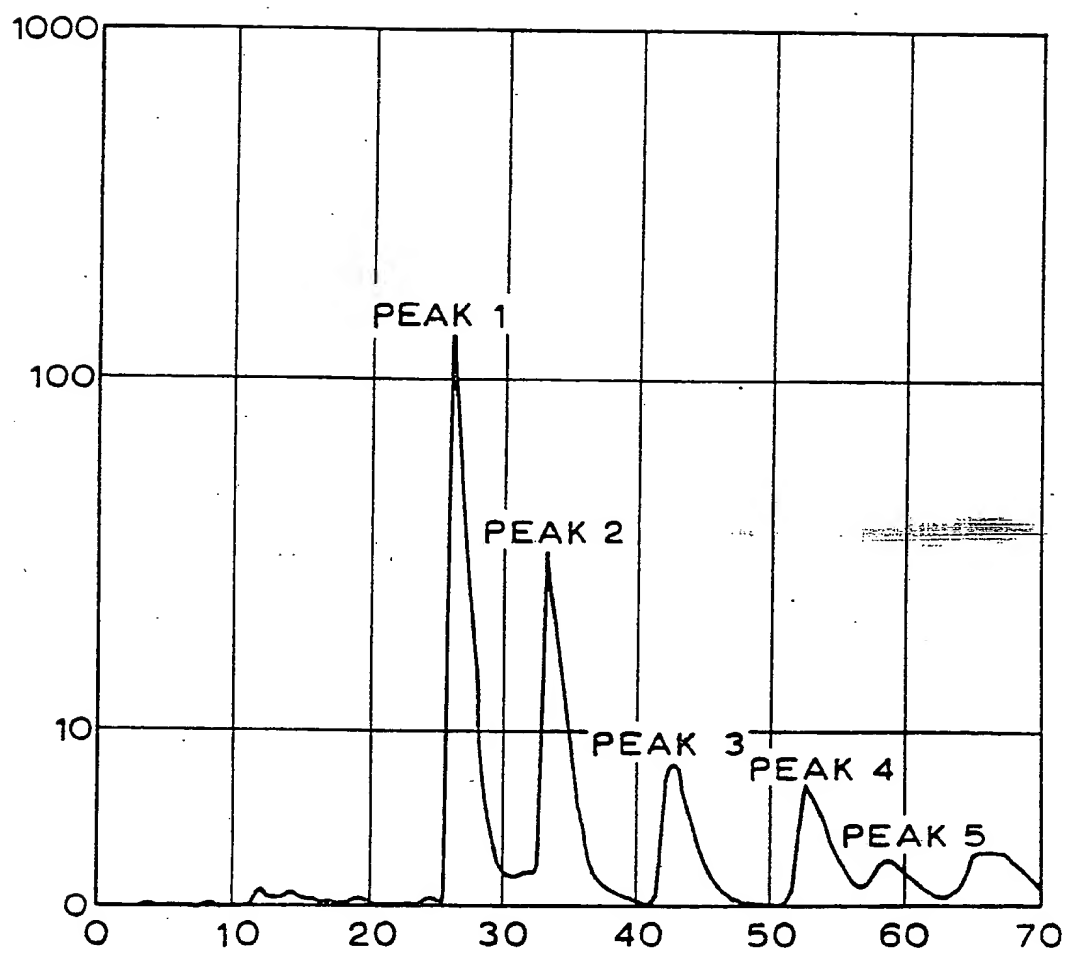


FIG. 2

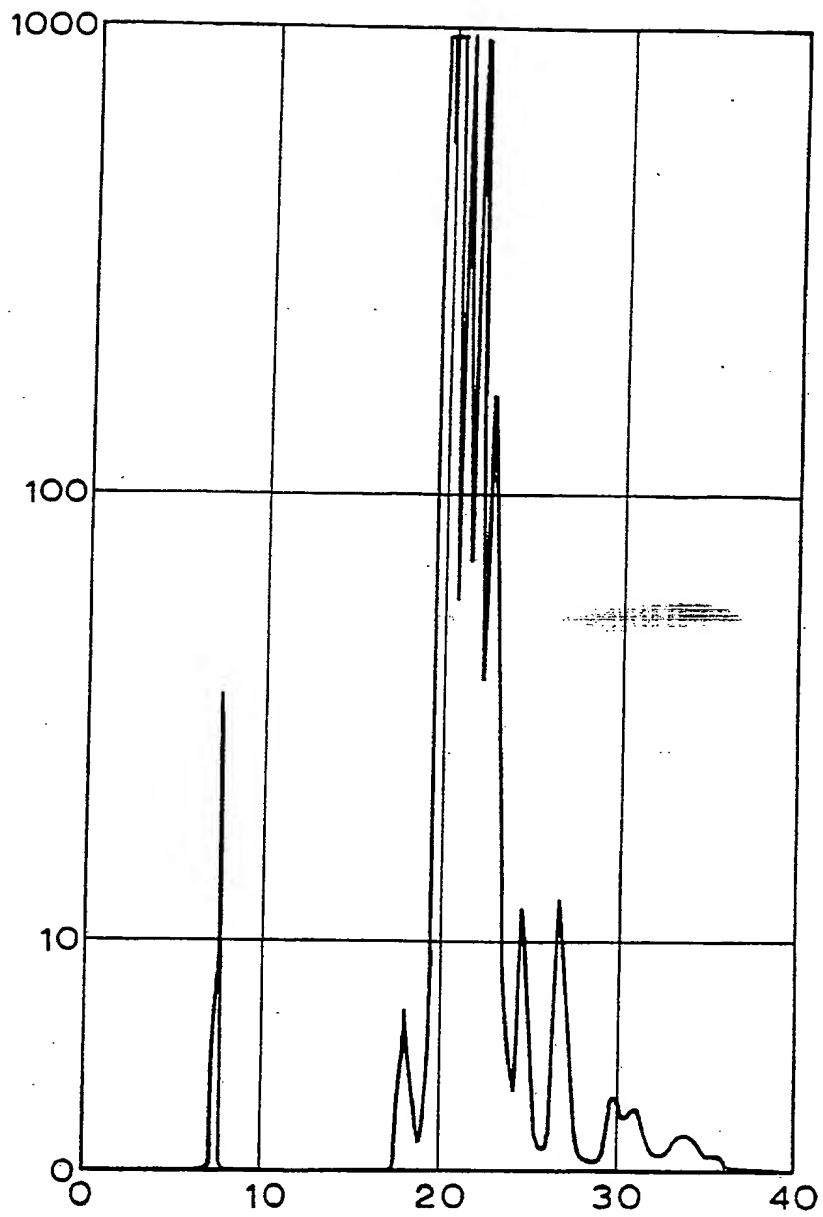


FIG. 3

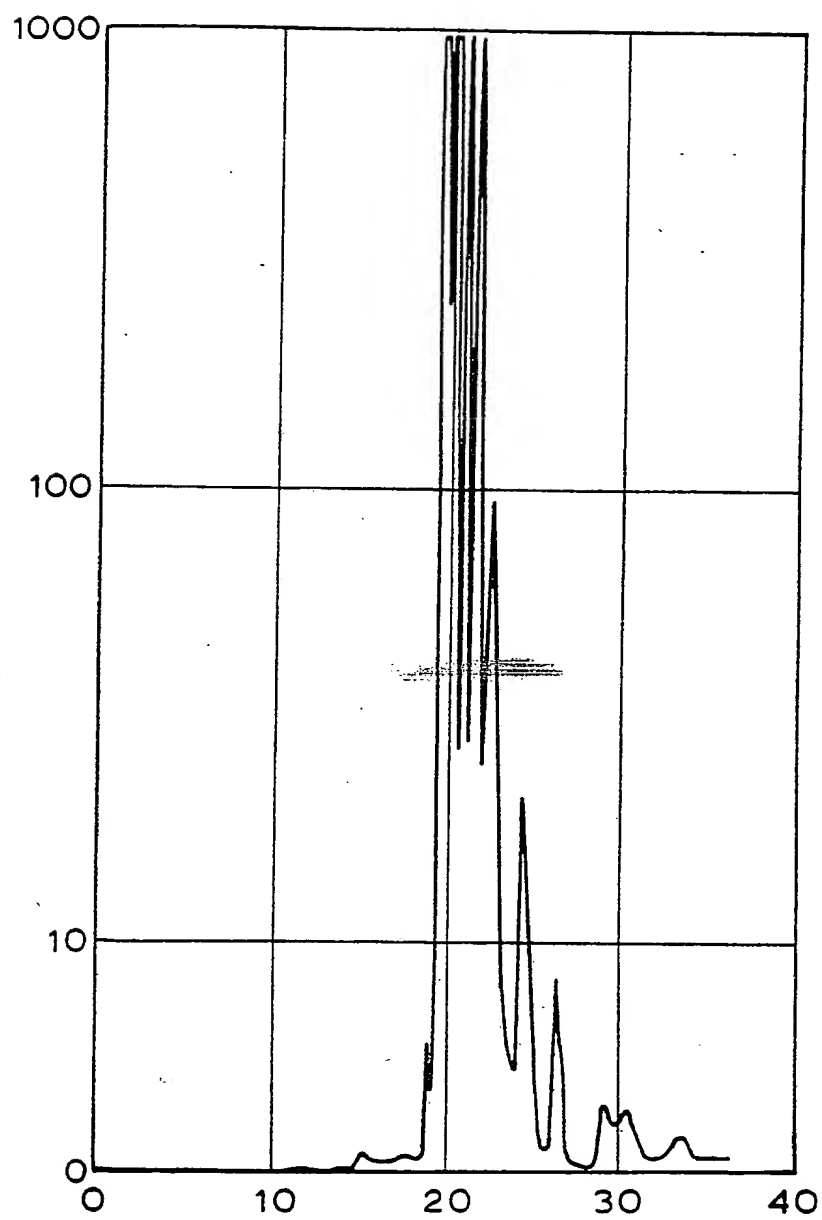


FIG. 4

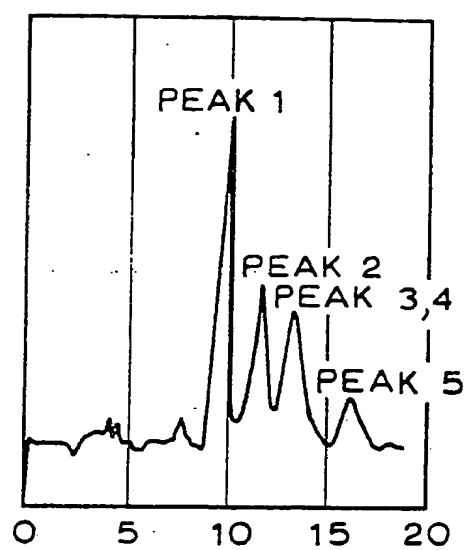


FIG. 5A

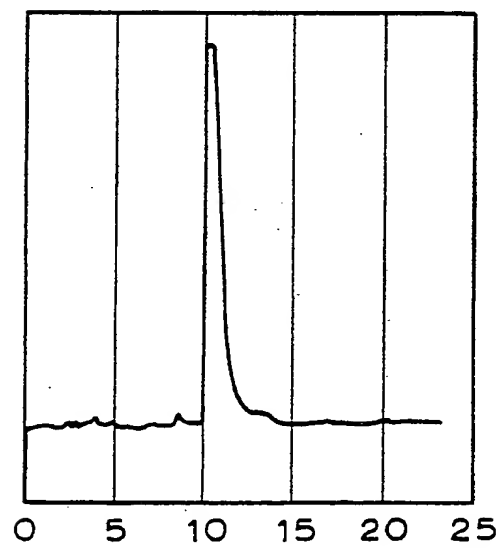


FIG. 5B

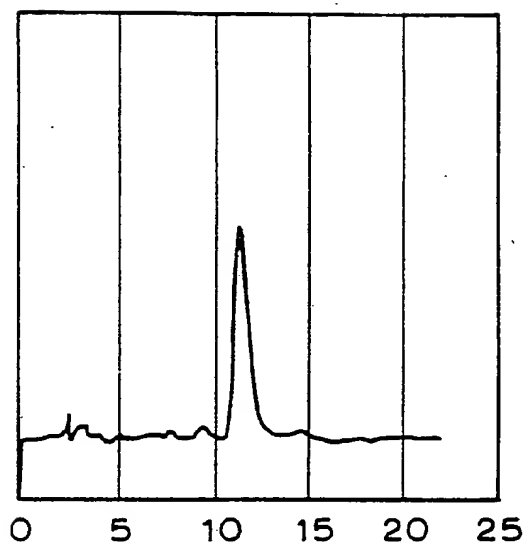


FIG. 5C

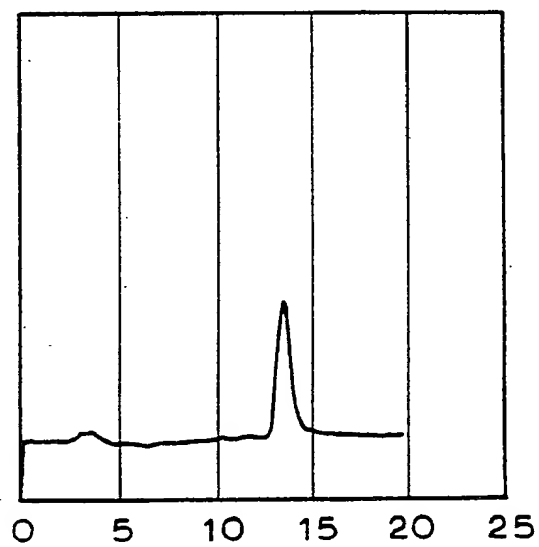


FIG. 5D

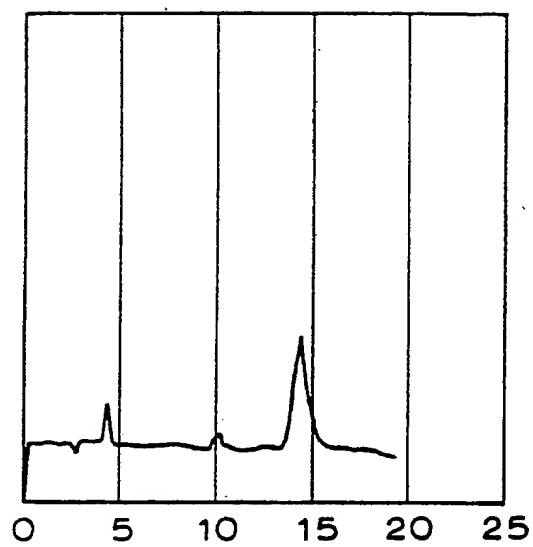


FIG. 5E

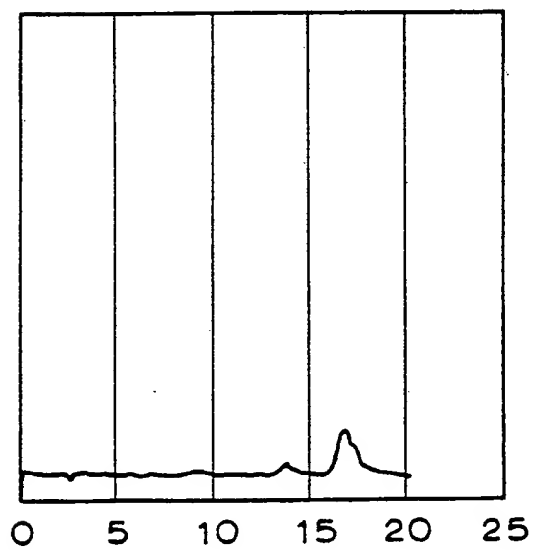


FIG. 5F

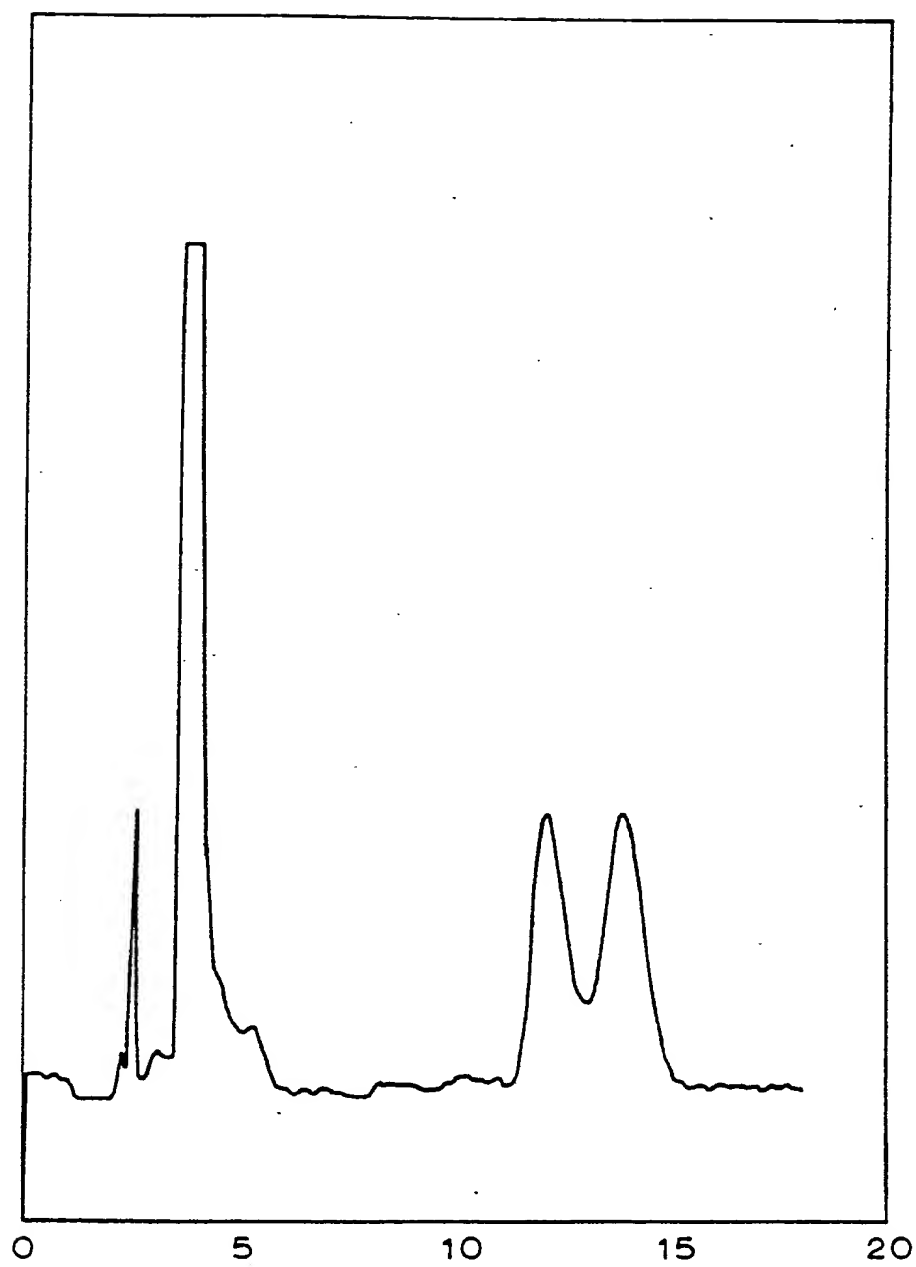


FIG. 6

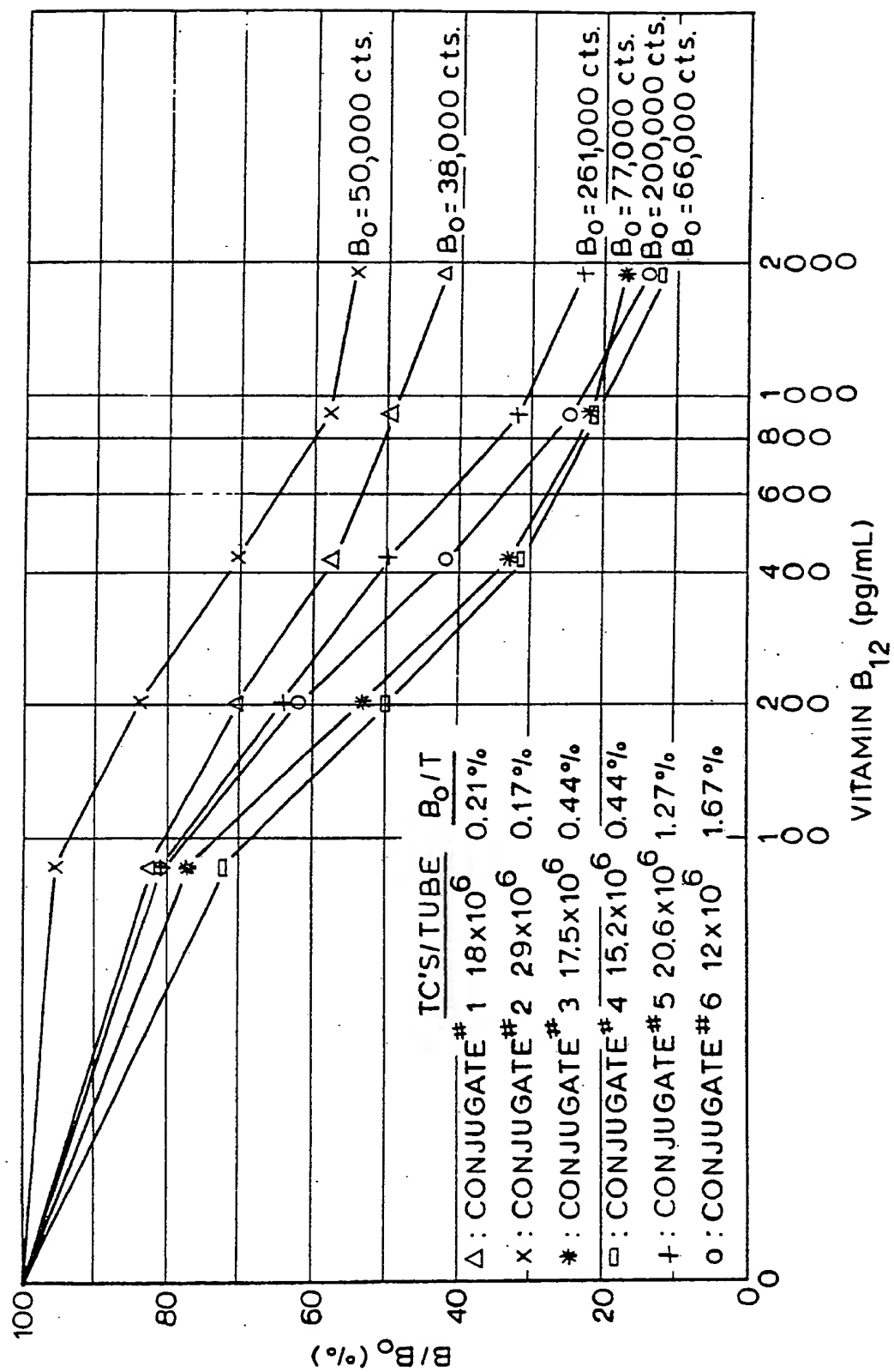


FIG. 7

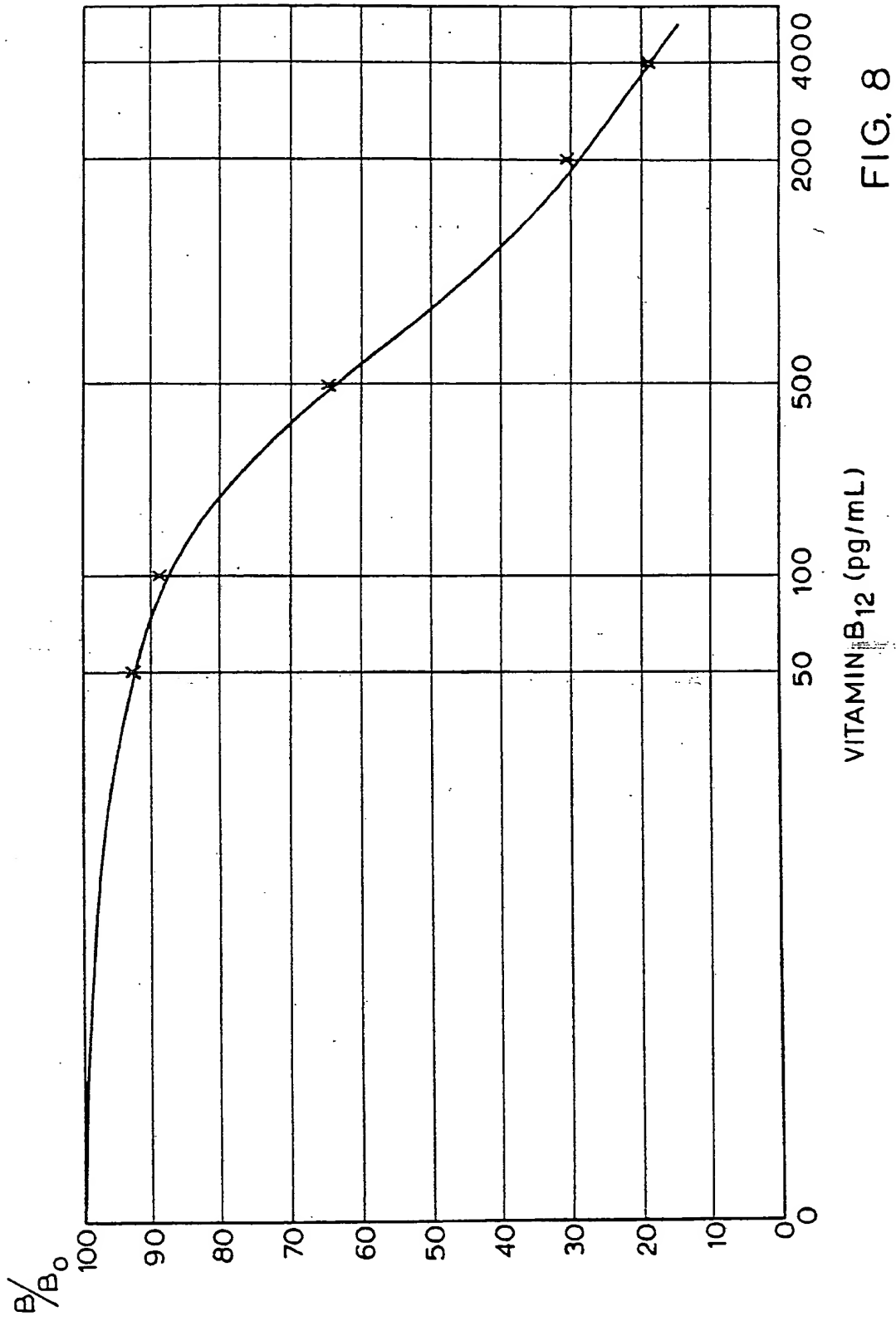


FIG. 8

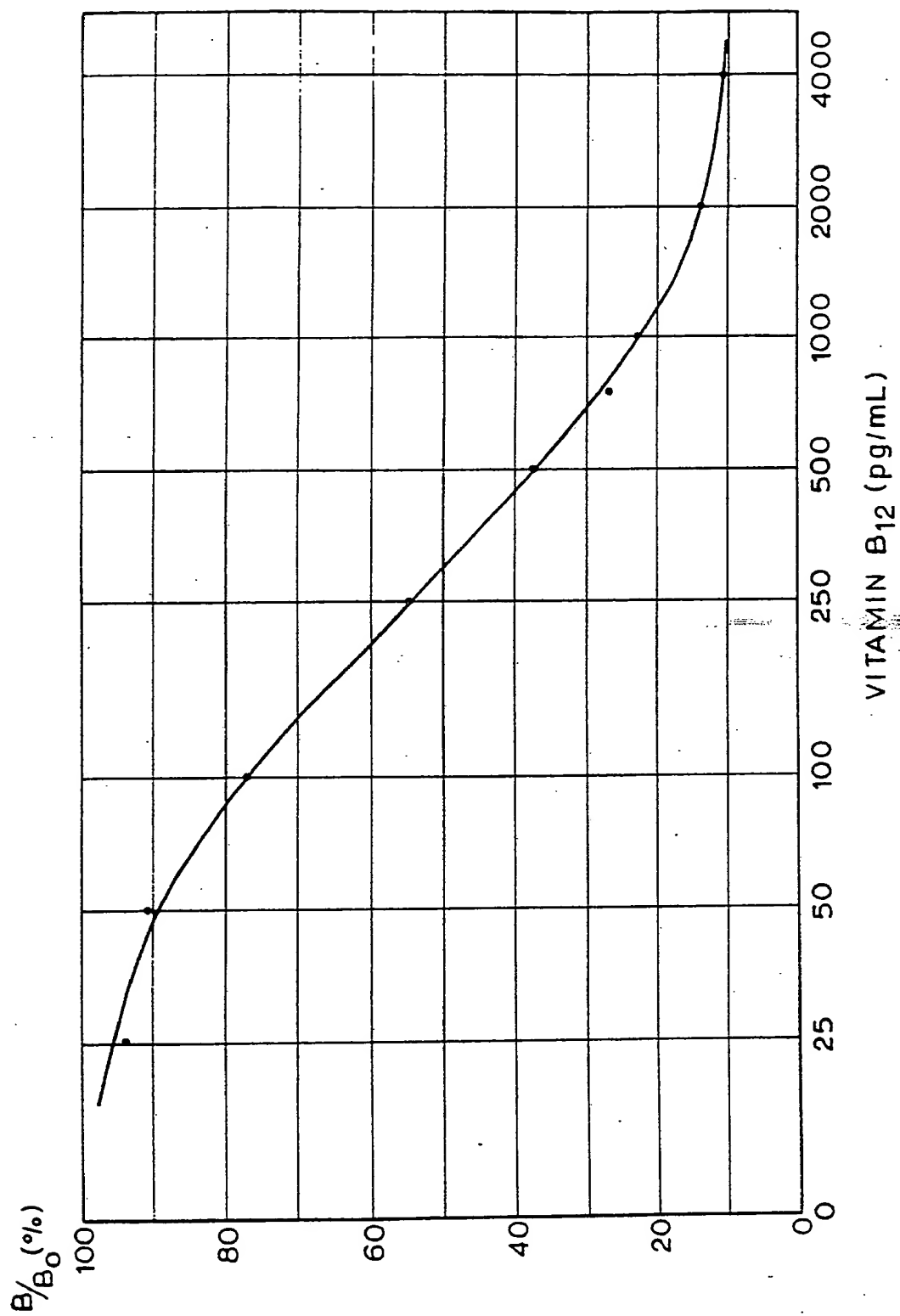


FIG. 9

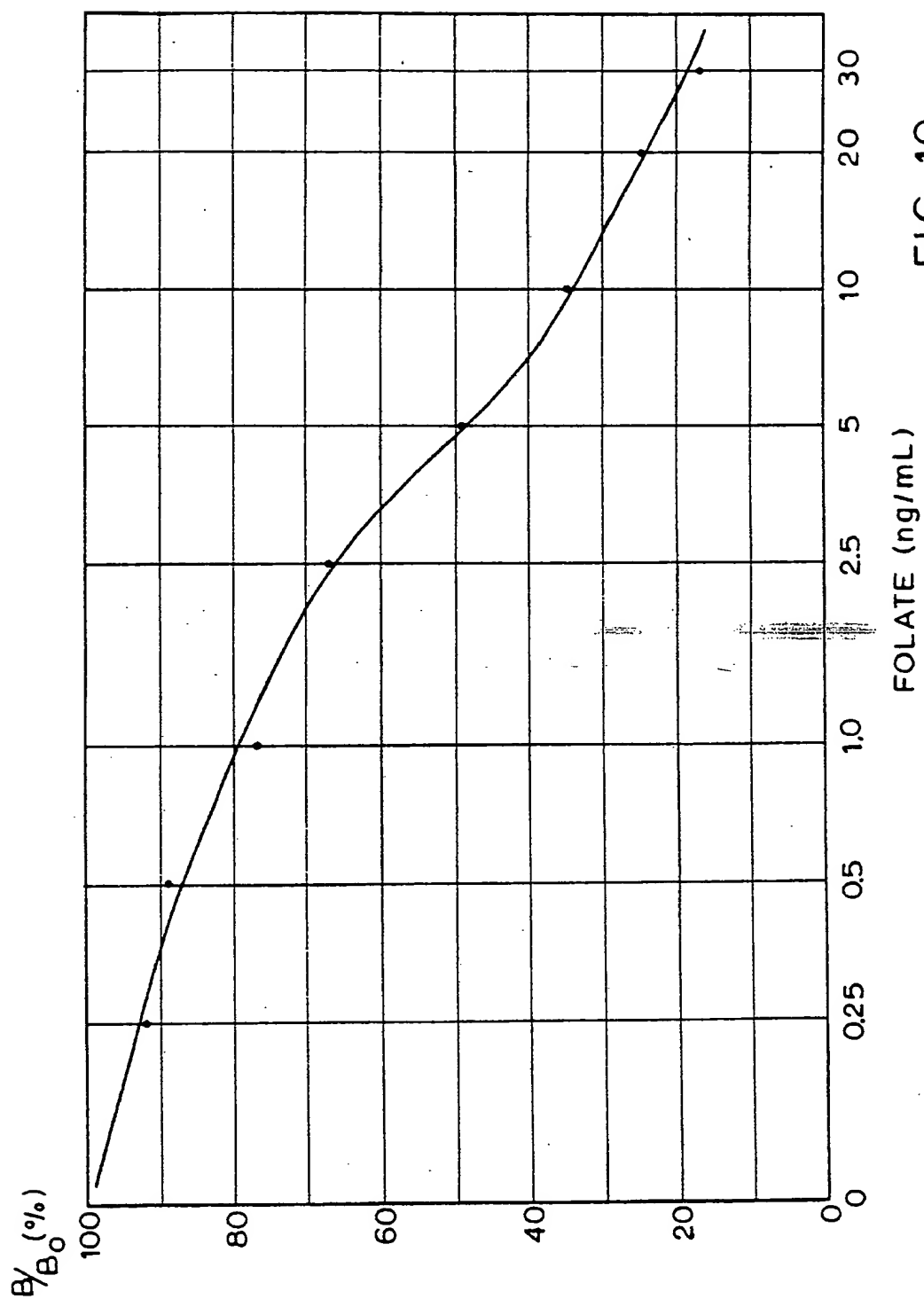


FIG. 10

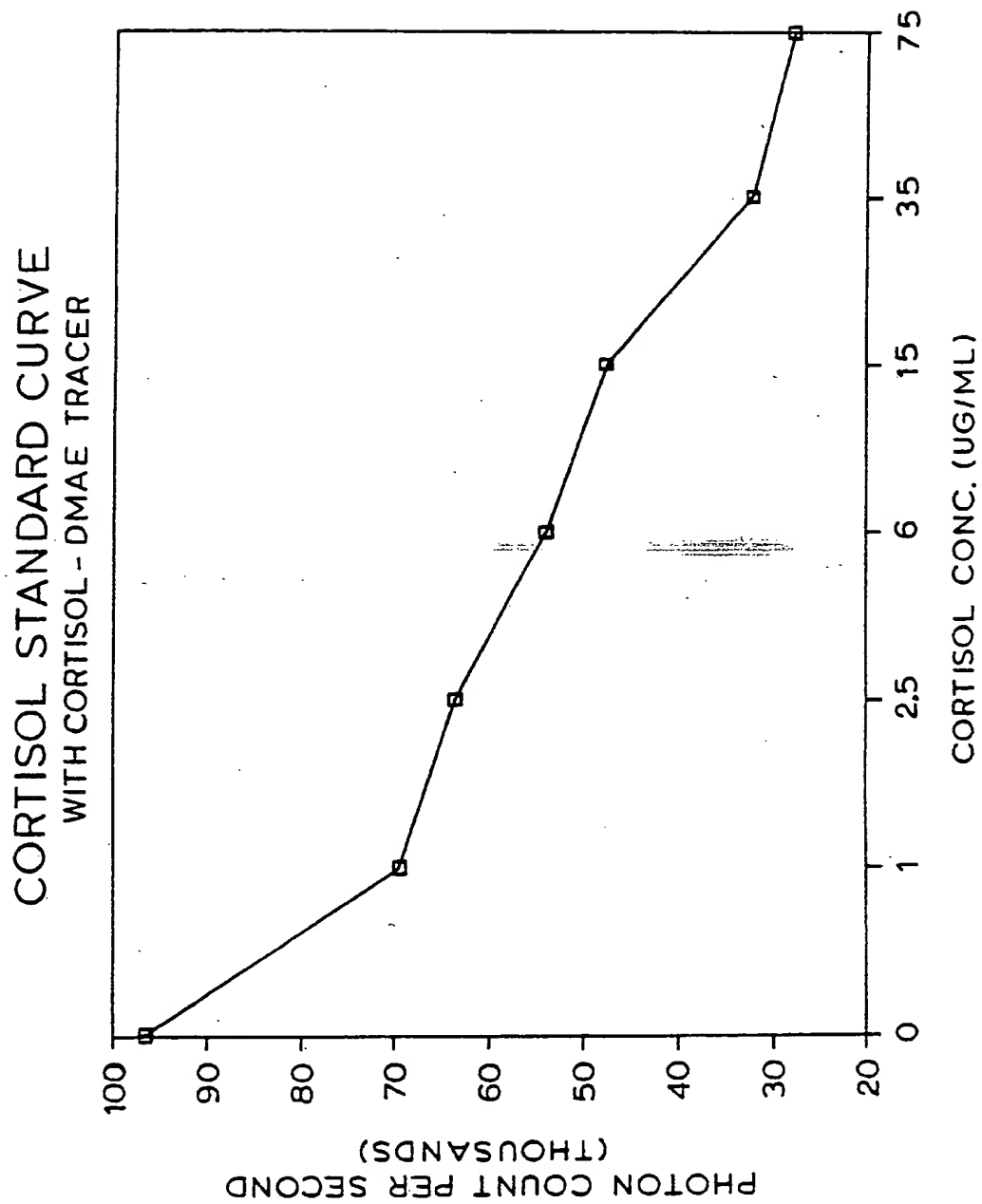


FIG. 11

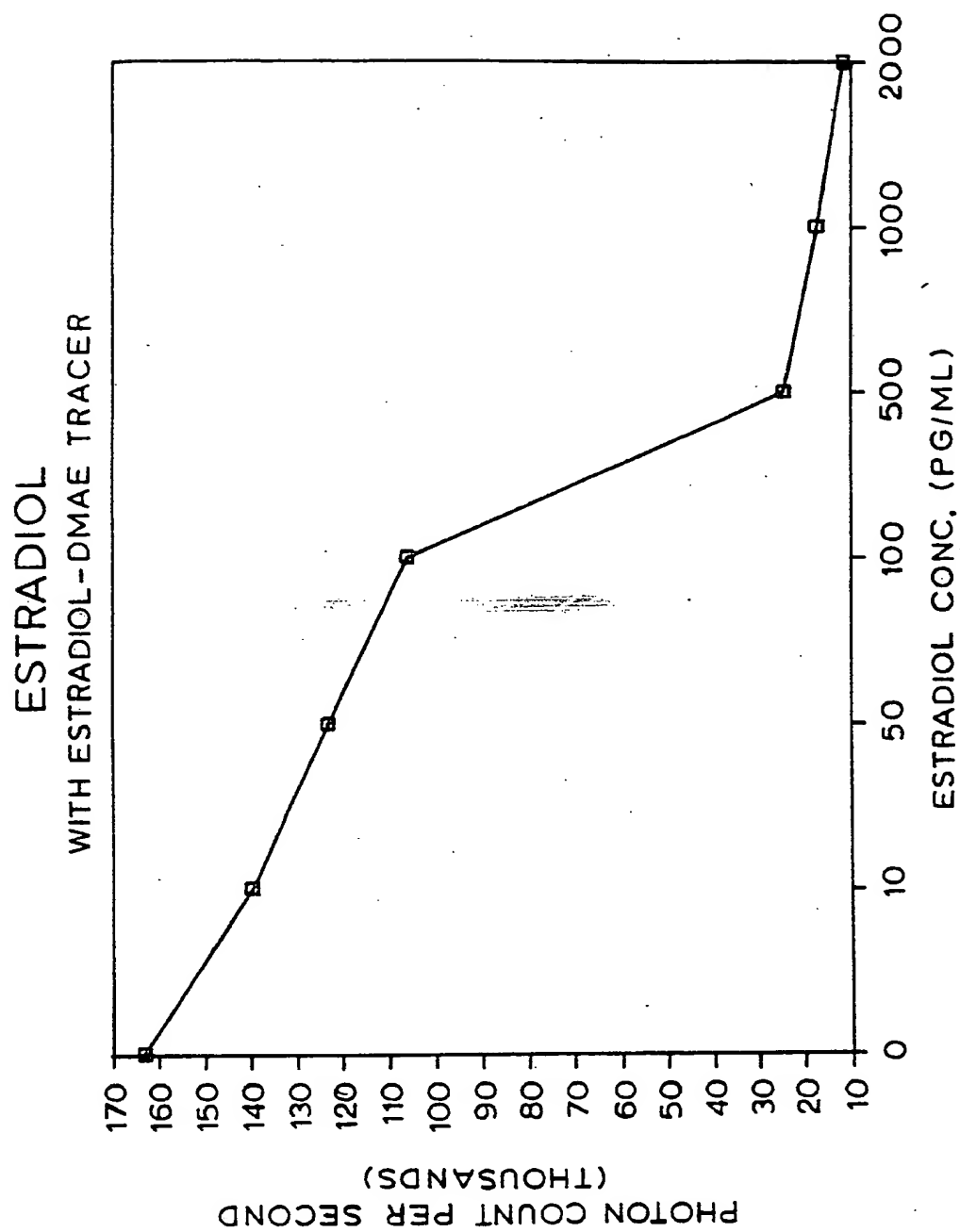


FIG. 12

TXB2 STANDARD CURVE WITH TXB2-DMAE TRACER

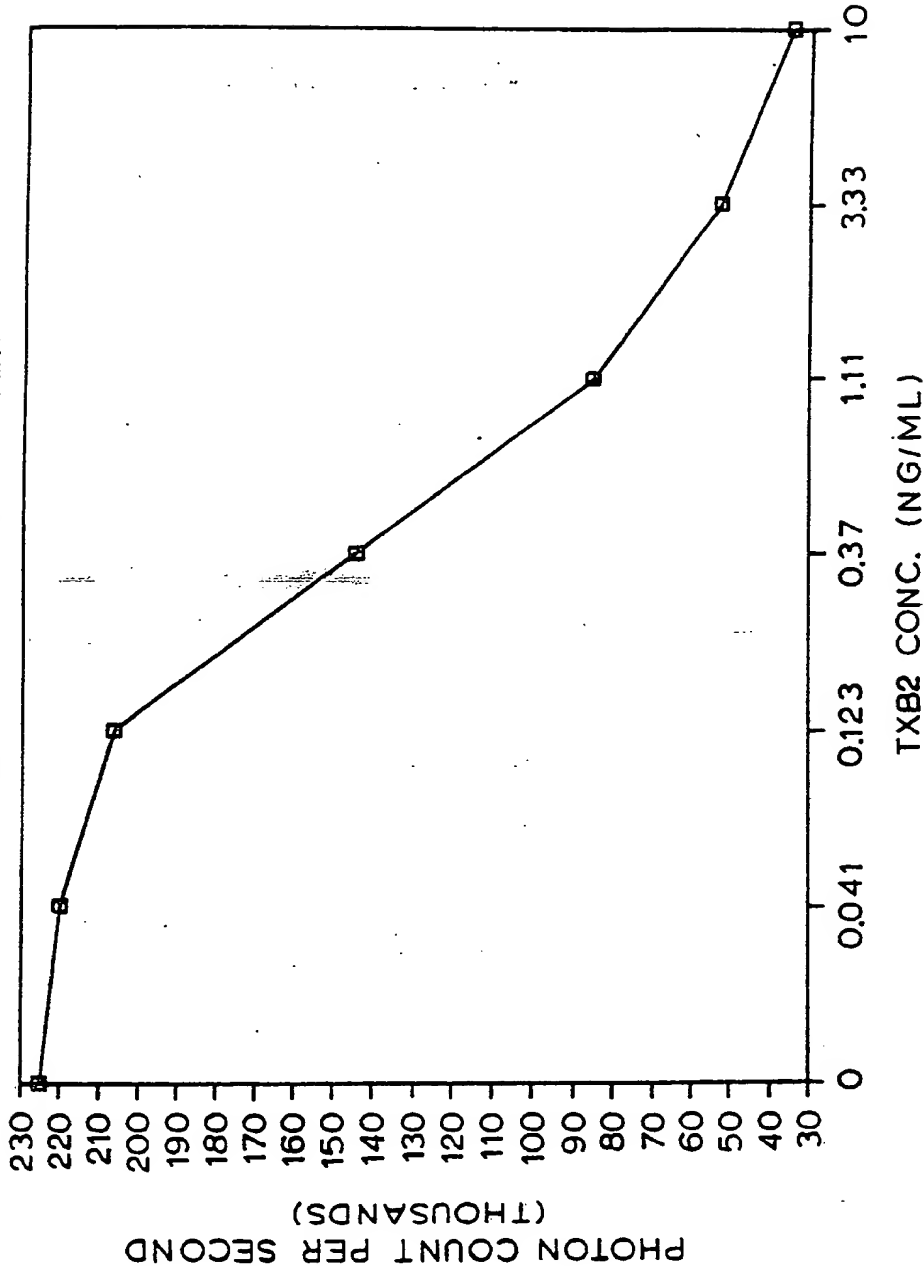


FIG. 13

(19)



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(54) Nucleophilic polysubstituted aryl acridinium esters, conjugates, production and uses thereof.

(57) Certain nucleophilic polysubstituted aryl acridinium esters are disclosed, as well as conjugates thereof, which are useful in biological assays, including assays for the determination of vitamin B₁₂, folate, cortisol, estradiol and thromboxane B₂.

EP 0 361 817 A3



European Patent
Office

EUROPEAN SEARCH REPORT

Application Number

EP 89 30 9705

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5)
D,A	EP-A-0 082 636 (THE WELSH NATIONAL SCHOOL OF MEDICINES) * Claims * ---	1,5,6	C 07 D 219/04 G 01 N 33/533 G 01 N 33/82 G 01 N 33/74 G 01 N 33/88
D,A	EP-A-0 263 657 (CIBA CORNING DIAGNOSTICS CORP.) * Claims * ---	1,5,6 /	
A	CHEMICAL ABSTRACTS, vol. 97, no. 11, 13th September 1982, pages 111-112, abstract no. 85407p, Columbus, Ohio, US; A. PATEL et al.: "The preparation and properties of a chemiluminescent derivative of 173-estradiol", & SERONO SYMP. PUBL. RAVEN PRESS 1982, 1(LUMIN. ASSAYS: PERSPECT. ENDOCRINOL. CLIN. CHEM.), 181-9 * Abstract * ---	1,5,6,9 ,15	
A	CLINICAL CHEMISTRY, vol. 29, no. 8, August 1983, pages 1474-1479, Washington, DC, US; I. WEEKS et al.: "Acridinium esters as high-specific-activity labels in immunoassay" * The whole article * ---	1,5,6	TECHNICAL FIELDS SEARCHED (Int. Cl.5) C 07 D 219/00 G 01 N 33/00
D,A	CLINICAL CHEMISTRY, vol. 31, no. 10, October 1985, pages 1664-1668, Washington, DC, US; A.P. RICHARDSON et al.: "Chemiluminescence immunoassay of plasma progesterone, with progesterone-acridinium ester used as the labeled antigen" * The whole article * -----	1,5,6	
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 05-07-1990	Examiner HENRY J.C.
CATEGORY OF CITED DOCUMENTS X: particularly relevant if taken alone Y: particularly relevant if combined with another document of the same category A: technological background O: non-written disclosure P: intermediate document T: theory or principle underlying the invention E: earlier patent document, but published on, or after the filing date D: document cited in the application L: document cited for other reasons &: member of the same patent family, corresponding document			